

Full Length Research Paper

Acute toxicity and behavioral response of freshwater fish, *Mystus vittatus* exposed to pulp mill effluent

A. Mishra¹, C. P. M. Tripathi², A. K. Dwivedi³ and V. K. Dubey^{3*}

¹Aquatic Toxicology Division, IITR Lucknow, Uttar Pradesh, India.

²D. D. U. Gorakhpur University, Gorakhpur, India.

³National Bureau of Fish Genetic Resources, Canal Ring Road, PO, Dilkusha, Lucknow- 26002, Uttar Pradesh, India.

Accepted 3 April, 2011

The present study deals with the acute toxicity (LC₅₀ evaluation) of paper mill effluent to a freshwater fish, *Mystus vittatus*. A monthly record covering all the spawning season (pre-spawning phase, spawning phase and post-spawning phase) was evaluated at different exposure periods (24-, 48-, 72- and 96-h) using the whole paper mill effluent. Annual variation in LC₅₀ values in relation to annual breeding cycle and water temperature were also taken into consideration. A well marked variation in the LC₅₀ values in different exposure periods as well as in various months of the spawning phases of the test fish were observed.

Key words: Acute toxicity, paper mill effluent, annual variation, spawning phases.

INTRODUCTION

The Bagridae family of fish is the richest and most important of the teleostei class and its members are distributed throughout the world (Day, 1878). In the Bagridae family, the fish *Mystus vittatus* (Smith, 1945) is economically important and distributed in the semitemporal freshwater system of South India (Perennou and Santharam, 1990).

Under laboratory conditions, toxicity testing procedures (mortality studies LC₅₀ estimates) may provide information regarding the harmfulness of industrial stress for aquatic animals, including fishes (Marier, 1973). In the acute toxicity of contaminants in static bioassays, the use of 96-h, LC₅₀ has been widely recommended as a preliminary step in toxicological studies on fishes (McLeay, 1976; Whittle and Flood, 1977; USEPA, 1973, 2005; APHA, 1998, 2005; Chapman, 2000; Ali and Sree-Krishnan, 2001; ASTM, 2002; Parrott et al., 2006; Moreira et al., 2008). The static bioassay procedures to study the toxicity of pulp and papermill effluent to variety

of fishes have been reported like the Cohosalmon, *Onchorhynchus kisutch* (McLeay, 1973, 1975, 1976, 1977; Gordon and McLeay, 1977; McLeay and Howard, 1977; McLeay and Gordon, 1977; McLeay and Brown, 1979) the rainbow trout, *Salmo gairdneri* (McLeay, 1976; Gordon and McLeay, 1977; Whittle and Flood, 1977; McLeay and Gordon, 1978; Couillard et al., 1988) and other fishes (Gordon and Servizi, 1974; Davis, 1976; Walden, 1976; Hewitt et al., 2006).

The paper mill, when it is playing directly influx to the streams and rivers, without previous treatment of effluents it can change the native fish or biota. Therefore, in the present study, the acute toxicity of paper mill effluent has been studied with regards to the mortality of *M. vittatus*. The 24-, 48-, 72- and 96-h, LC₅₀ values were determined during the different phases of the reproductive cycle of the fish, and the correlation of LC₅₀ values with environmental temperature has also been established.

The main objective of this study is to evaluate the acute toxicity of paper mill effluent on the behavioral responses of freshwater fish *M. vittatus* in the context of annual breeding cycle and water temperature.

*Corresponding author. E-mail: vineet_dubey26@yahoo.co.in.

Table 1. Characteristics of WPME (whole paper mill effluent), Magahar, Santkabeer Nagar (U.P.) India.

Characteristics	Variable constituents through the year (mean values)			Yearly average \pm S. E.
	January-April	May-August	September-December	
Color	Dark brownish	Dark brownish	Dark brownish	-
Sodium, Na ⁺ (mg/L)	350	320	351	340 \pm 12.2
Cl ⁻ (mg/L)	420	450	425.7	431.9 \pm 11.3
SO ₄ (mg/L)	1.12	5.8	3.5	3.5 \pm 1.7
Nitrate (mg/L)	7.6	7.3	7.2	7.4 \pm 0.1
Total nitrogen (mg/L)	1.7	6.3	3.8	3.9 \pm 1.6
PO ₄ (mg/L)	0.77	0.52	0.69	0.7 \pm 0.1
pH	7.3	7.3	7.4	7.3 \pm 0.04
Temp (°C)	23.5	28	26.5	26 \pm 1.6
Suspended solid (mg/L)	5021	4643	4256	4640 \pm 270
Dissolved solid (mg/L)	1111	1222	1013	1115 \pm 74
Total solid (mg/L)	6132	5865	5269	5755 \pm 312
BOD (mg/L)	552	538	498	526 \pm 19
COD (mg/L)	2379	2551	2326	2418 \pm 83
Fe (mg/L)	9.6	13.5	10.4	11.2 \pm 1.5
Mg (mg/L)	1.9	1.65	1.28	1.6 \pm 0.2
K (mg/L)	8.6	6.4	4.4	6.5 \pm 1.5
Cu (mg/l)	0.11	0.14	ND	0.1 \pm 0.1
Total Cr (mg/L)	ND	0.077	0.072	0.07 \pm 0.03
Mn (mg/L)	0.34	0.071	0.53	0.31 \pm 0.12
Co (mg/L)	-	0.001	0.0012	0.001 \pm 0.002
Cd (mg/L)	0.025	0.016	0.018	0.02 \pm 0.004
Zn (mg/L)	0.06	0.08	0.106	0.08 \pm 0.021

Data based on samples taken during the morning shift of the normal course of mill operation at 8 am.

MATERIALS AND METHODS

The freshwater teleost fish, *M. vittatus* were collected from a local freshwater river Ami of Sant Kabeer nagar district (U. P). They were transported to laboratory and washed with KMNO₄ solution (1 mg/L) for 5 min and then transferred to the acclimation tank. The fishes having an average body length of 7.6 \pm 0.18 cm, and body weight of 73 \pm 0.23 g. were selected for the present study.

The fishes were acclimatized in laboratory conditions for 3 to 4 weeks at room temperature (22 to 30°C) using dechlorinated tap water. The medium of the acclimation tanks was changed daily. The dissolved oxygen content of the medium was maintained at 60% to 100% by regular aeration. The whole paper mill effluent was collected in polyethylene container of 10 to 20 L capacity during the morning hours. The effluent samples were taken from two places, that is, the point where the effluent comes out of the mill and the point where it enters the collecting tank and the mixed unfiltered samples were used within 24 h of collection. The effluent samples were chemically analyzed per month and the characteristics are summarized in Table 1.

Acute toxicity of effluent on test fish was measured in terms of LC₀, LC₅₀ and LC₁₀₀. Static bioassay procedures, as outlined by the USEPA (2005) were followed. A minimum of 8 concentrations of effluent and 20 animals were used for each concentration. The control set had the same number of fish kept in normal, dechlorinated tapwater. No food was provided to either the control or the test fishes during the period of the toxicity experiments. Experiments were carried out up to 96-h, and the observations were recorded after every 24-h. Experiments were conducted every

month, and LC₅₀ values for 24-, 48-, 72- and 96-h, were determined. The tested concentrations of effluent and the observed percentage of the fish mortality were subjected to linear regression for determining the LC₅₀ values for the different exposure periods. The yearly average LC₅₀ values were calculated, and the 95% confidence interval determined for this mean.

RESULTS

The mortality of test fish exposed to paper mill effluent is summarized in Table 2.

Yearly average LC₅₀

The yearly average LC₅₀ value was found to be 60.04 \pm 2.13; for 24 h, 53.6 \pm 2.16, for 48 h; 48.6 \pm 2.1, for 72 h, and 44.4 \pm 2.2, for 96 h exposure periods (Table 2).

Annual variation in LC₀ and LC₁₀₀

The lowest LC₀ and LC₁₀₀ values were recorded in the spawning phase of the fish, whereas the values were the highest in the post spawning phase (October to January)

Table 2. Seasonal variation of WPME toxicity (LC₅₀) to *M. vittatus* in different spawning phases.

Spawning phases	Month	Water temperature (°C)	LC ₅₀ % (v/v)			
			24 h	48 h	72 h	96 h
Pre spawning phase	February	23.5	64.5	54.0	49.5	45.6
	March	24.0	62.5	54.5	50.5	45.7
	April	25.0	60.4	53.5	46.05	41.5
	May	27.0	54.0	48.5	43.5	42.5
	Mean			60.35	52.6	47.8
Spawning phase	June	29.0	50.0	42.55	38.0	35.5
	July	28.0	50.5	42.55	38.0	37.0
	August	28.5	53.0	46.0	41.5	38.0
	September	27.0	57.25	53.75	49.0	44.25
	Mean			52.7	46.2	41.6
Post-spawning phase	October	26.5	64.5	59.5	50.5	46.0
	November	25.0	68.5	62.5	58.0	51.5
	December	23.0	71.75	64.0	59.0	53.0
	January	22.0	68.25	61.75	58.0	53.0
	Mean			68.2	62.0	56.4
Yearly average ±S. E.		25.7±0.7	60.4±2.13	53.6±2.16	48.6±2.1	44.4±2.2
95% Confidence interval		-	65.1-55.7	58.3-48.8	53.2-43.9	49.2-39.6

of the annual reproductive cycle. Values observed in three phases are recorded in Table 3.

Annual variation in LC₅₀

A well marked variation was recorded in the LC₅₀ values for the different months of the year. The values for the spawning phase were generally lower than those of other phases (Table 2).

Variation in relation to water temperature

The annual variation in LC₅₀ values exhibited an inverse relationship with water temperature. The LC₅₀ value was generally recorded to be higher in the colder months of the year and lower in the hotter months. Maximum LC₅₀ values were observed in the month of December (water temperature 23°C), while lowest in the month of July (water temperature 28°C.).

The Cr and Cd contents of test effluent were found to be much higher; the total content is 0.497 mg/L for Cr and 0.0073 mg/L for Cd. Regarding Cu, the maximum permissible concentration is 0.006 ppm or alternatively 0.1 times the LC₅₀ 96 h value. In case of the presently used papermill effluent, the Cu content was found to be 0.083 mg/L, which is about 2.5 times its estimated LC₅₀ 96 h value (that is, estimated concentration present in the LC₅₀ 96 h test effluent).

The matter suspended within the papermill effluent used in the present study could also be contributing to the mortality of the *M. vittatus* individuals maintained in this effluent. The concentration of suspended matter in WRPBILE ranged from 4256 to 5021 mg/L.

Behavioral responses

The surfacing behavior as well as the rate of opercular movement of the fishes was observed to be increased within an hour of the commencement of the toxicity experiments. With the continuation of the exposure, the fishes progressively became sluggish and lethargic. The majority of them became completely inactive after 72 h of exposure to 60 to 70% of test effluent, and all died after 96 h of exposure to 70% effluent. Prior to their death in the contaminated medium, they showed abnormal swimming and the loss of equilibrium.

DISCUSSION

The stressed fishes were found to be the most sensitive to effluent during the spawning phases of the reproductive cycle. Increased sensitivity under pollution stress during the spawning season has also been reported in pacific *herrings* exposed to benzene toxicity (Korn et al., 1976). Higher temperature nature has been shown to increase the toxicity of pulp and papermill

Table 3. Seasonal variation of WRPBILE toxicity (LC₀ and LC₁₀₀) to *M. vittatus* in different spawning phases.

Spawning phase	Months	Water temperature (°C)	LC ₅₀ % (v/v)							
			24 h		48 h		72 h		96 h	
			LC ₀	LC ₁₀₀	LC ₀	LC ₁₀₀	LC ₀	LC ₁₀₀	LC ₀	LC ₁₀₀
Pre-spawning phase	February	23.5	30.0	85.0	30.0	80.0	25.0	75.0	25.0	70.0
	March	24.0	30.0	80.0	30.0	75.0	25.0	75.0	25.0	70.0
	April	25.0	25.0	80.0	25.0	75.0	20.0	70.0	20.0	65.0
	May	27.0	25.0	75.0	20.0	70.0	20.0	65.0	20.0	60.0
	Mean		27.5	80.0	26.3	75.0	22.5	71.3	22.5	66.3
Spawning phase	June	29.0	20.0	75.0	20.0	70.0	20.0	65.0	20.0	60.0
	July	28.0	20.0	70.0	20.0	65.0	15.0	60.0	15.0	55.0
	August	28.5	25.0	75.0	20.0	70.0	20.0	65.0	15.0	60.0
	September	27.0	30.0	80.0	25.0	75.0	25.0	75.0	20.0	70.0
	Mean		23.7	75.0	21.3	70.0	20.0	66.3	17.5	61.3
Post-spawning phase	October	26.5	30.0	80.0	30.0	75.0	25.0	75.0	25.0	70.0
	November	25.0	40.0	85.0	35.0	80.0	35.0	80.0	30.0	75.0
	December	23.0	40.0	90.0	40.0	85.0	35.0	80.0	30.0	75.0
	January	22.0	40.0	85.0	40.0	80.0	35.0	75.0	30.0	70.0
	Mean		37.5	85.0	36.3	80.0	32.5	77.5	28.8	72.5
Yearly average ±S.E.		25.7±0.7	29.6±2.2	80.0±2.1	28.0±1.8	75.0±2.1	25.0±1.8	71.7±1.9	22.9±1.5	66.7±1.9
95% confidence Interval		-	34.4 -24.7	84.6 -75.3	31.9-24.0	79.6-70.3	28.9-21.0	75.8-67.5	26.2-19.6	70.8-62.5

effluent for fishes like *O. kisutch* and *S. gairdneri* (McLeay, 1976; Gordon and McLeay, 1977). Increase in temperature reduces the solubility of oxygen in water and hence, could raise the metabolic rate (oxygen demand) of the fish, thus limiting the oxygen carrying capacity of the blood (Gordon and McLeay, 1977). But in the present study, the LC₅₀ played the higher toxicity in the colder months, when the temperatures were very low. The result indicates that toxicity can occur independent of the high or low temperatures. Simultaneously, the higher BOD (Biological oxygen demand) and COD (Chemical oxygen

demand) could also contribute to the mortality (Polak and Palmer, 1977; Whittle and Flood, 1977; McLeay et al., 1979a, b; McLeay and Brown, 1979).

It is known that many pollutants, such as heavy metals, like Mg, Cu and Zn, and industrial wastes become more harmful at higher water temperatures. In view of this, it may be presumed that the toxic metals, especially Cu and Zn, present in the tested effluent would acquire greater toxicity during the warmer months of the year and thus contribute to the overall increased mortality during these months in *M. vittatus*. As

regard the contribution of the different constituents of effluent to *M. vittatus* mortality, the content of many of these constituents is noticeably higher than the upper limits considered as safe for fish health (Wedemeyer, 1976). For example, among metals, the upper limit for Cr is 0.03 ppm, and that for Cd 0.0004 ppm. The total suspended solids, absorbable organic halogens and also certain other toxic compounds like wood extractives, present in pulp and papermill waste water are also known to affect fishes (Sprague and McLeese, 1968; Kelso, 1977; Hewitt et al., 2006; Parrott et al., 2006). The concentration of the suspended

matter in WRPBILE is much above the TSS concentration considered (Wedemeyer, 1976) optimum for fishes (that is, 80 to 100 ppm), and falls within that range of suspended matter of papermill effluents (that is, 530 to 17900 mg/L approximately) which has earlier been considered injurious to fishes.

The opercular movement of the dying fishes became extremely slow and they were seen to secrete thick coat of mucus profusely around their opercular region. Similar types of increased opercular movement, erratic and rapid movements have been observed by Kumar and Gopal (2001) in *Channa punctatus* exposed to distillery effluent. Clotfelter et al., (2006) have observed aggressive behaviour in fighting fish, increased surface breathing and opercular movement in the stressed *M. vittatus* would point a sustained respiratory discomfort in the toxic WPME (whole paper mill effluent) (Lloyd, 1961). The stressed fishes were observed to secrete mucus around their opercular region which has been considered to be a symptom of the inflammatory reaction of gill towards the pollutants (Durve and Jain, 1980 ; Pandey and Pandey, 1988), while the abnormal swimming and disturbed orientation has been considered as a generalized sign of diseases in fishes (Sindermann, 1970).

The kinds of behavioral faults in orientation and locomotion, as observed in the present study, can be related to the impairment of sensory organ systems particularly the mechano and chemo-receptor systems. Sensory organs like lateral line, olfactory organs and membranous labyrinth helps the fishes in maintaining harmony with their environments and also control their vital behaviors (Gardner, 1975). Hence, any impairment of these organs would produce behavioral faults in the fishes. Therefore, the behavioral changes, particularly those concerned with respiratory insufficiency, observed in *M. vittatus* exposed to mill effluent, might be contributing to the mortality in these stressed fishes.

ACKNOWLEDGEMENTS

The authors are grateful to Prof. C. P. M. Tripathi, HOD Department of Zoology, D. D. U., Gorakhpur University and Dr. Krishna Gopal, for encouragement and valuable support.

REFERENCES

- Ali M, Sreekrishnan TR (2001). Aquatic toxicity from pulp and paper mill effluents: A review. *Adv. Environ. Res.*, 5: 175-196. doi: 10.1016/S1093-0191(00)00055-1.
- American Public Health Association (APHA). American Water Work Association (AWWA) and Water Pollution Control Federation (WPCF) (1998). Standard method for the examination of water and waste water. 20th ed. Washington, DC.
- APHA (2005). American Public Health Association. Standard methods for examination of water including bottom sediments and sludges. Standard Methods, (19th ed.), p. 874.
- ASTM (American Society for Testing and Materials) (2002). Standard guide for conducting acute toxicity tests on test materials with fishes, macro vertebrates and amphibians. E729/96. In Annual Book of ASTM standards, ASTM, Philadelphia, PA., 11.05: 179–200.
- Chapman PM (2000). Whole effluent toxicity test-usefulness, level of protection and risk assessment. *Environ. Toxicol. Chem.*, 19: 3-13. doi:10.1897/1551-5028(2000).019<0003:WETTUL>2.3 Co:2.
- Couillard CM, Berman RA, Panisset JC (1988). Histopathology of rainbow trout exposed to a bleached kraft pulp paper mill effluent. *Environ. Contam. Toxicol.*, 17: 319-323.
- Clotfelter D, Ethane C, Alison R (2006). Behavioural changes in fish exposed to phytoestrogens. *Environ. Pollut.*, 144: 833-839.
- Davis JC (1976). Progress in sublethal effect studies with kraft pulp papermill effluent and salmonids. *J. Fish. Res. Board Canada* 33: 2031-2035.
- Day F (1878). The Fishes of India. William Dawson, London. 1.2: 210-215.
- Durve VS, Jain SM (1980). Toxicity of distillery effluent to the cyprinid weed fish *Rasbora daniconius* (Ham). *Acta. Hydrochim.*, 8(4): 329-336.
- Gardner GR (1975). Chemically induced lesions in estuarine or marine teleosts. In: The Pathology of fishes (Eds: Ribelin, W.E. and Migaki, G.M.). Wisconsin Press, Madison, pp. 657-693.
- Gordon MR, McLeay DJ (1977). Sealed-Jar bioassays for pulp mill effluent toxicity: Effects of fish species and temperature. *J. Fish Res. Board, Canada*, 30: 1389-1396.
- Gordon RW, Servizi JA (1974). Acute toxicity and detoxification of kraft pulp mill effluent. *Int. Pac. Salmon Fish. Comm. Rep.*, p. 31.
- Hewitt LM, Parrott JL, McMaster ME (2006). A decade of research on the environmental impacts of pulp and papermill effluents in Canada: Sources and characteristics of bioactive substances. *J. Toxicol. Environ. Health*, 9:341-356. doi:10.1080/15287390500195976.
- Kelso JRM (1977). Density, distribution and movement of Nipigon Bay fishes in relation to a pulp and paper mill effluent. *J. Fish. Res. Board Canada*, 34: 879-885.
- Korn S, Hirsch N, Struhsaaker JN (1976). Uptake, distribution and depuration of 14C-benzene in Northern anchovy, *Encrallus mordax*, and striped bass, *Monero saxatilis*. *Fish. Bull.*, 74: 545.
- Kumar S, Gopal K (2001). Impact of distillery effluent on physiological consequences in the fresh water teleost *Channa punctatus*. *Bull. Environ. Contam. Toxicol.*, 66: 617-622.
- Lloyd R (1961). Effect of dissolved oxygen concentrations on the toxicity of several poisons to rainbow trout (*Salmo gairdneri*. *Rich.*). *J. Exp. Biol.*, 38: 447.
- Marier JR (1973). The effects of pulp and paper-wastes on aquatic life with particular attention to fish and bioassay procedures for assessment of harmful effects. *NRC (Nat. Res. Counc. Com.) Publ. No. 73-3(ES)*, p. 33.
- McLeay DJ (1973). Effects of a 12-h and 24-day exposure to Kraft pulp mill effluent on the blood and tissue of Juvenile cohosalmon (*Onchorhynchus kisutch*). *J. Fish. Res. Board Canada*, 30: 395-400.
- McLeay DJ (1975). Sensitivity of Blood cell counts in Juvenile cohosalmon (*Onchorhynchus kisutch*) to stressors including sublethal concentrations of pulp mill effluent and Zinc. *J. Fish. Res. Board Canada*, 32: 2357-2364.
- McLeay DJ (1976). A rapid method for measuring the acute toxicity of pulp mill effluents and other toxicants to salmoid fish at ambient room temperature. *J. Fish. Res. Board Canada*, 33: 1303-1311.
- McLeay DJ (1977). Development of a blood sugar bioassay for rapid measuring stressful levels of pulp effluent to salmoid fish. *J. Fish. Res. Board Canada*, 34: 477-485.
- McLeay DJ, Brown DA (1979). Stress and chronic effects of untreated and treated bleached kraft pulp mill effluent on the biochemistry and stamina of Juvenile cohosalmon (*Onchorhynchus kisutch*). *J. Fish. Res. Board Canada*, 36: 1049-1059.
- McLeay DJ, Gordon MR (1977). Leucocrit: A simple hematological technique for measuring acute stress in salmonid fish including stressful concentration of pulp mill effluent. *J. Fish. Res. Board Canada*, 34: 2164-2175.
- McLeay DJ, Gordon MR (1978). Effect of seasonal photoperiod on acute toxic responses of Juvenile rainbowtrout (*Salmo gairdneri*) to pulp mill effluent. *J. Fish. Res. Board Canada*, 32: 1388-1392.
- McLeay DJ, Howard TE (1977). Comparison of rapid bioassay

- procedures for measuring toxic effects of bleached kraft mill effluent to fish. Proc. 3rd Aquatic Toxicity workshop, Halifax, N.S., Nov. 2-3, 1976. Env. Prot. Serv. Tech. Report No. EPS-5_AR-77-1, Halifax, Canada, pp. 141-155.
- McLeay DJ, Walden CC, Munro JR (1979a). Influence of dilution water on the toxicity of kraft pulp and papermill effluent including mechanisms of effect. *Water Res.*, 13: 151-158.
- McLeay DJ, Walden CC, Munro JR (1979b). Effect of pH on toxicity of kraft pulp and papermill effluent to salmonid fish in fresh and sea water. *Water Res.*, 13: 249-254.
- Moreira-Santos M, Donato C, Lopes I, Ribeiro R (2008). Avoidance tests with small fish: determination of the median avoidance concentration and of the lowest-observed-effect gradient. *Environ. Toxicol. Chem.*, 27: 1576-1582. doi:10.1897/07-094.1.
- Pandey RK, Pandey SK (1988). Tolerance measurement and histopathological observations on gills of *Mystus. M. vittatus* under the toxic stress of the fertilizer NPK. *Acta hydrochim. Hydrobiol.* 17(5): 597-601.
- Parrott JL, McMaster ME, Hewitt LM (2006). A decades of research on the environmental impacts of pulp and papermill effluents in Canada: Development and application of fish bioassays. *J. Toxicol. Environ. Health*, 9: 297-317 doi:10.1080/15287390500195752.
- Perennou C, Santharam V (1990). Anthropological survey of some wetlands in south-east India. *J. Bombay Nat. Hist. Soc.*, 87: 354-363.
- Polak J, Palmer MD (1977). Concentration pattern of chemical constituents in papermill effluent plume: Dynamics and model. *J. Fish. Res. Board Canada*, 34: 805-816.
- Sindermann CJ (1970). *Principal diseases of fish and shellfish*. Academic Press, London/New York, p. 369
- Sprague JB, McLeese DW (1968). Different toxic mechanisms in kraft pulp mill effluent for two aquatic animals. *Water Res.*, 2: 761-765.
- Walden CC (1976). The toxicity of pulp and papermill effluents and corresponding measurement procedures. *Water Res.*, 10: 639-664.
- Whittle DM, Flood KW (1977). Assessment of the acute toxicity, growth impairment, and flesh taining potential of a bleached Kraft mill effluent on rainbow trout *Salmo gairdneri*. *J. Fish Res. Board Canada*, 34: 869-878.
- U.S. Environmental Protection Agency, (USEPA) (1973). *Proposed Criteria for Water Quality*. US Environmental Protection Agency, Washington DC, USA, Vol. 1.
- United States Environmental Protection Agency (USEPA) (2005). *Aquatic Toxicity Information Retrieve AQUIRE aquatic toxicology database*. Available:/www.epa.gov/ecotox/accessed:August 2003.
- Wedemeyer GA (1976). Physiological response of juvenile coho salmon (*Oncorhynchus kisutch*) and rainbow trout (*Salmo gairdneri*) to handling and crowding stress in intensive fish culture. *J. Fish. Res. Board Can.*, 33: 2699-2702.