

Full Length Research Paper

Exploration of the neurotoxicity of ciprofloxacin or gatifloxacin single dose in rat cortex and hippocampus

Nadia Mohamed Said Arafa^{1*}, Sayed M. Rawi² and Sara Abdullah Mubarak³

¹Department of Biology, Faculty of Science, Jazan University, KSA & National Organization for Drug Control and Research, Department of Physiology, Egypt.

²Department of Biology, Faculty of Sciences and Arts, Khulais, Jeddah University, Saudi Arabia.

³Department of Dairy Lab, Public Authority of Agriculture & Fish Resources (PAAFR), State of Kuwait.

Received 15 November, 2014; Accepted 29 January, 2015

The study aimed to evaluate the neurotoxicity of ciprofloxacin (Cip) or gatifloxacin (Gati) single oral dose in male albino rats weighing (100 ± 20 g) grouped as control-administered water, ciprofloxacin (80 mg/kg) and gatifloxacin (32 mg/kg) each of 12 rats. The frontal cortex of both groups revealed decrease in glutamate, GABA, taurine, histidine and serotonin levels and elevation of aspartate, glycin and serine and AChE activities. While noradrenaline and dopamine levels reduced significantly in Gati group, noradrenaline increased significantly in Cip group. Hippocampus of either Cip or Gati group's results revealed elevation of all detected amino acids and monoamines except the reduction of glutamate, aspartate and dopamine in Cip group. In the meantime, AChE activities significantly reduced in both treatments. Serum results showed elevation of glucose in both treated groups. The histological examination of Gati brain tissue showed neuronal degeneration in the cerebral cortex and congestion in the blood vessels and capillaries in hippocampus tissue without histopathological alteration observed in Cip group tissue. Overall, the data showed the effect of the quinolones single dose towards hyperglycemia and shift in balance of neurotransmitters and acetylcholinesterase as well as the histopathological alterations in the tested brain areas.

Key words: Ciprofloxacin, gatifloxacin, cortex, hippocampus, neurotransmitters, glucose.

INTRODUCTION

Gatifloxacin is one of the broad-spectrum fluoroquinolones available and approved by the US food and drug administration (FDA) in December 1999. Ever since its release in the market, there have been numerous reports implicating gatifloxacin as a cause of

hypoglycemia and hyperglycemia. This prompted Bristol-Meyer Squibb Co. to list diabetes mellitus as a contraindication to gatifloxacin use in the US product labeling and Health Canada to issue an advisory against the use of gatifloxacin in patients with diabetes (Jose et

*Corresponding author. E-mail: nadianeuro@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](https://creativecommons.org/licenses/by/4.0/)

al., 2007). Gatifloxacin showed to be equivalent to ciprofloxacin for the treatment of acute uncomplicated lower urinary tract infections (Naber et al., 2004). In extensive *in vivo* and *in vitro* experiments performed in an attempt to explain the central nervous system (CNS) side effects of quinolones sometimes observed under therapeutic conditions, they are like dizziness, restlessness, tremor, insomnia, hallucinations, convulsions, anxiety and depression. However, the molecular target or receptor for such effects is still not exactly known. Extensive toxicological and biochemical experiments were performed to explain the CNS effects observed under therapeutic conditions (Akahane et al., 1993; De Sarro et al., 1999, De Sarro and De sarro 2001).

Seizure activity is associated with a wide range of local biochemical changes, affecting various neurotransmitters (monoamines, amino acids) (Freitas et al., 2004; Cavalheiro et al., 2006). Cortex and hippocampus areas appeared to be important in the expression of early convulsive seizures (Kelly et al., 1999; Ang et al., 2006) in addition to the important functional association between cortical regions and the hippocampus in seizure propagation (Kelly et al., 2002) and suggested playing a role in inducing convulsions by quinolones (Motomura et al., 1991). The US Food and Drug Administration (FDA) Safety Announcement (8-15-2013) has recently issued a warning about fluoroquinolone antibacterial drugs; serious side effect of peripheral neuropathy may occur soon after these drugs are taken and may be permanent.

The study designed using single oral dose of the tested quinolones to explore its neurotoxicity as the single dose in accordance with previous studies where it was used for treatment (Loo et al., 1985), randomized controlled trials (Boy et al., 2004; Kaushik et al., 2010; Heidari Bateni et al., 2014) and its prophylactic activity (Terzi et al., 2005; Alborzi et al., 2008). The study aims to ascertain the effect of oral single dose of either Cip or Gati in male albino rat on the concentrations of amino acid and monoamine neurotransmitters and acetylcholinesterase activities in the frontal cortex and hippocampus brain areas and the histopathological examination of both areas, in addition to the determination of serum glucose level.

MATERIALS AND METHODS

Experimental animals

This study carried was out on thirty-six adult male albino rats (*Rattus norvegicus*) with average body weight of range 100 ± 20 g obtained from the Egyptian Institution of Serum and Vaccine (Helwan). The experiment was conducted in the Department of Physiology in National Organization for Drug Control and Research (NODCAR). The male albino rats were housed in iron mesh cages

with seven rats each. Clean sawdust was used to keep the animals dry and clean throughout the experimental period. The experimental animals were allowed acclimating under the laboratory conditions two weeks before the beginning of the experiments. The animals were kept under controlled temperature of 21°C and 12 h light/12 h dark cycle throughout the course of experiment. A commercial pelleted diet was used during the experiment and allowed water *ad libitum*.

Drugs

Ciprofloxacin (Cipro) ($C_{17}H_{18}FN_3O_3 \cdot HCl \cdot H_2O$), manufactured by Bayer healthcare pharmaceuticals, Ciprofloxacin hydrochloride tablets and Gatifloxacin (TEQUIN) ($C_{19}H_{22}FN_3O_4 \cdot 1.5 H_2O$) manufactured by Bristol-Myers Squibb Company. The antibiotics were administered by gastric intubation technique and doses calculated equivalent to the human therapeutic dose according to the Guidance for Industry and Reviewers (2002).

Experimental design

Animals were divided into three groups using random selection; the first group (n = 12 rats) was administered 2 ml of distilled water, the second group (Cip) was administered 80 mg/100 g body weight ciprofloxacin dissolved in 2 ml water. The Gati-treated rat groups (n = 48 rats) was administered 32 mg/100 g body weight gatifloxacin dissolved in 2 ml water. Animals were sacrificed after 12 h from dose administration by rapid decapitation. Blood samples were collected and sera separated for assessment of glucose using the BioAssay Systems' glucose assay kit (QuantiChrom™ Glucose Assay Kit). The brains were dissected out quickly, weighed and cleaned. Four brains from each treated group were served for the histopathological examination according to Banchroft et al. (1996) and the rest eight brains for the biochemical analysis. The frontal cortex and hippocampus areas were separated and divided into two halves; the first half was served for acetylcholinesterase activity assay according to the modification of Ellman et al. (1961) method as described by Gorun et al. (1978). The second half was homogenized in 75% high performance liquid chromatography (HPLC) methanol (1/10 weight/volume) using a homogenizer surrounded with an ice jacket. The homogenates were used for the determination of the brain contents of amino acids using the precolumn PTC derivatization technique according to method of Henrikson and Meredith (1984) and monoamines neurotransmitters according to method described by Pagel et al. (2000).

Statistical analysis

Reported values were represented as means \pm SE. Statistical analysis was evaluated by one-way ANOVA. Once a significant F-test was obtained, least significance difference (LSD) comparisons was performed to assess the significance of differences among various groups using statistical processor system support "SPSS" for Windows software, Release 20.0 (SPSS, Chicago, IL).

RESULTS

The data as presented in Figure 1 as percentage change from control about frontal cortex showed decrease in

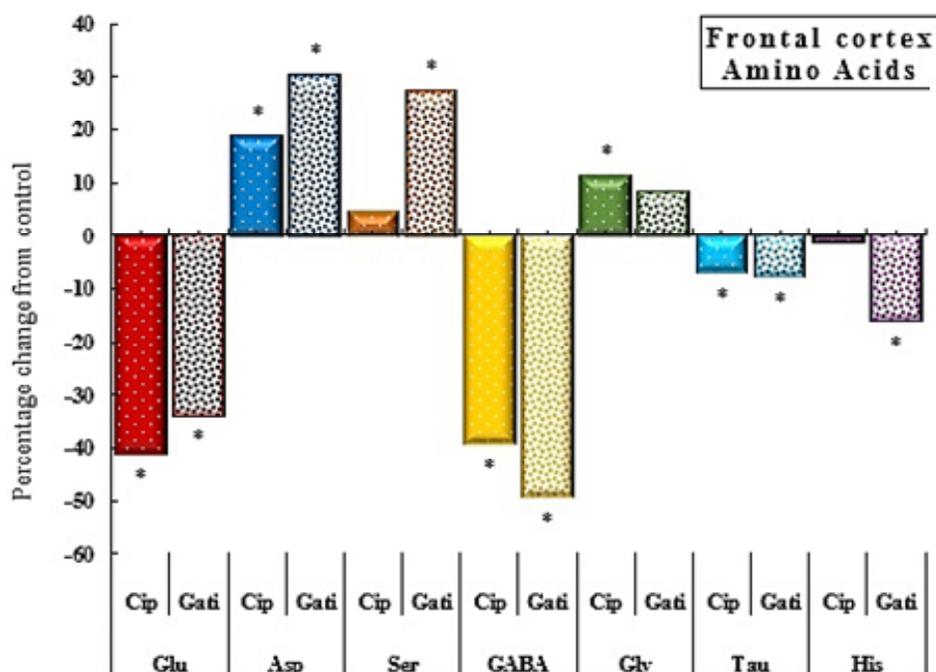


Figure 1. Percentage change from control of amino acids in cortices of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

levels of glutamic acid, GABA, taurine and histidine and increase in aspartic, glycine and serine levels post administration of either ciprofloxacin or gatifloxacin. In ciprofloxacin group, glutamic acid, GABA, taurine and histidine data as mean \pm SE is given as 5.32 ± 0.17 (-40.97%), 1.51 ± 0.05 (-39.02%), 1.86 ± 0.04 (-6.85%) and 1.20 ± 0.01 (-1.15%), respectively, while in gatifloxacin group, results are given as 5.99 ± 0.15 (-33.60%), 1.26 ± 0.003 (-49.21%), 1.85 ± 0.06 (-7.45%) and 1.03 ± 0.02 (-15.76%) different from control values 9.02 ± 0.25 , 2.47 ± 0.04 , 2.00 ± 0.05 and 1.22 ± 0.03 , respectively. Aspartic acid increased significantly after ciprofloxacin and gatifloxacin: 3.37 ± 0.06 (18.49%) and 2.84 ± 0.06 (30.10%), respectively, from control value 2.18 ± 0.04 . Serine level increased significantly after gatifloxacin: 0.60 ± 0.02 (27.23%) and not statistically different after ciprofloxacin administration: 0.49 ± 0.01 (4.47%), respectively, from control value 2.18 ± 0.04 . Glycine level increased significantly after ciprofloxacin: 1.90 ± 0.04 (10.97%) and not statistically different after gatifloxacin administration 1.85 ± 0.06 (8.05%) from control value 1.71 ± 0.06 .

The data represented in Figure 2 as percentage change from control about hippocampus showed no statistically different decrease in glutamic acid level and significant decrease in level of aspartic acid in

ciprofloxacin group recording 9.17 ± 0.20 (-6.72%) and 1.53 ± 0.06 (-20.67%), respectively from control values 9.83 ± 0.30 , 1.93 ± 0.05 , respectively. Meanwhile, serine, GABA, glycine and histidine levels increased significantly in ciprofloxacin group recording values, 0.25 ± 0.005 (11.89%), 2.46 ± 0.08 (27.67%), 1.14 ± 0.01 (13.91%) and 2.70 ± 0.09 (9.65%), respectively from control values 0.23 ± 0.01 , 1.92 ± 0.07 , 1.00 ± 0.02 and 2.47 ± 0.07 , respectively. Amino acids level in hippocampus of gatifloxacin group showed no statistically different increase in glutamic acid recording: 10.39 ± 0.26 (5.60%). Meanwhile, it showed significant increase in all detected amino acids recording values as: 1.98 ± 0.03 (2.37%), 0.28 ± 0.01 (22.03%), 3.17 ± 0.10 (64.59%), 1.44 ± 0.05 (44.04%), 3.62 ± 0.07 (7.75%) and 2.71 ± 0.06 (10.02%) for aspartic, serine, GABA, glycine, taurine and histidine, respectively.

Monoamines level and acetylcholinesterase activities recorded percentage change from control in frontal cortex and hippocampus of treated groups as well as serum glucose presented in Figure 3. In ciprofloxacin group, noradrenaline increased significantly in frontal cortex and hippocampus as mean \pm SE by 1.19 ± 0.03 (12.44%) and 0.65 ± 0.03 (15.60%), respectively. While in gatifloxacin group, noradrenaline decreased significantly in frontal cortex recording 0.78 ± 0.03 (-26.48%) and significantly

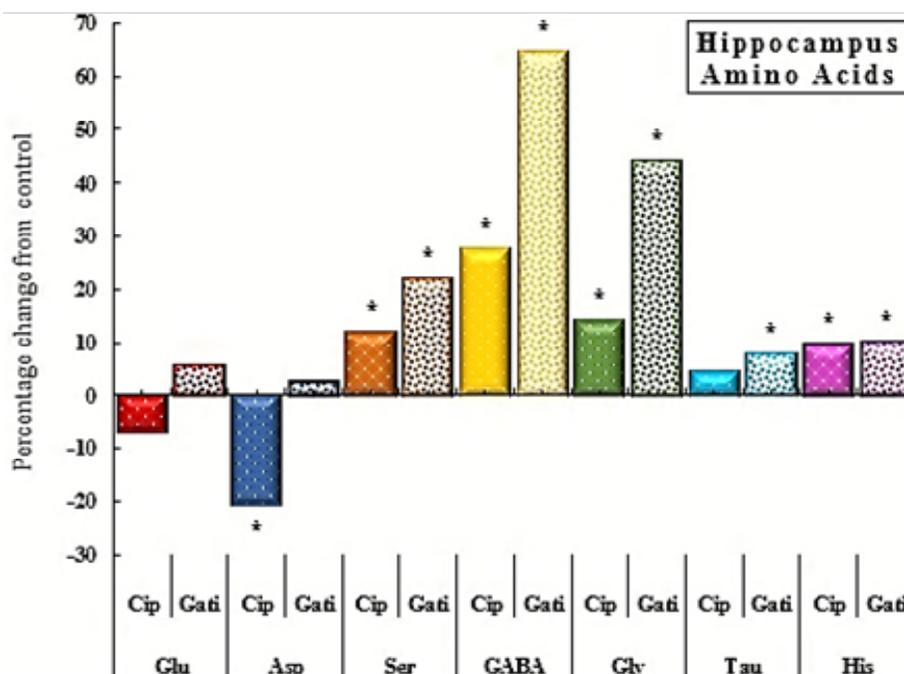


Figure 2. Percentage change from control of amino acids in hippocampi of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

increased in hippocampus 0.87 ± 0.01 (54.79) from control values 1.06 ± 0.004 and 0.56 ± 0.05 , respectively. Dopamine decreased significantly in frontal cortex of gatifloxacin group 2.95 ± 0.05 (12.21%) from control value 3.36 ± 0.11 . Serotonin level decreased significantly in cortical area of ciprofloxacin and gatifloxacin groups 0.07 ± 0.001 (-15.30%) and 0.48 ± 0.002 (-43.53%), respectively from control value 0.08 ± 0.003 . Meanwhile it increased significantly in hippocampus of gatifloxacin group 0.29 ± 0.004 (19.95%) from control value 0.25 ± 0.006 .

Acetylcholinesterase activities increased significantly in frontal cortex while it decreased significantly in hippocampus of both treatments recording in frontal cortex activities of 16.7 ± 0.61 (21.82%) and 15.64 ± 0.91 (17.84%) in ciprofloxacin and gatifloxacin groups, respectively from control value 13.27 ± 0.33 . While in hippocampus the data recorded was 15.47 ± 0.55 (-11.70%) and 16.23 ± 0.61 (-7.36%) in ciprofloxacin and gatifloxacin groups, respectively from control value 17.52 ± 0.25 . In addition, serum glucose level increased, recording 16.7 ± 0.61 (21.82%) and 121.88 ± 3.50 (40.69%) in ciprofloxacin and gatifloxacin groups, respectively from control value 86.63 ± 0.93 . With regards to the hispopathological examination, the response of cortex and hippocampus cells to Cip and Gati administration is represented in Figure 4A to D. Figure 4A and B showed

normal histology of cerebral cortex and hippocampus in control group. There was no histopathological alteration observed in hippocampus of Cip group in Figure 4C, while in Gati group there was neuronal degeneration in the cerebral cortex (Figure 4D) associated with congestion in the blood vessels and capillaries of the hippocampus (Figure 4E).

DISCUSSION

Fluoroquinolones had structural similarities to kynurenic acid and other similar compounds which are endogenous ligands of the glutamate receptor, which might suggest an interaction of quinolones with ligand-gated glutamate receptors as well (Schmuck et al., 1998), and may explain the effect on quinolones subjected groups. The excitatory potency of fluoroquinolones is based on activation of the N-Methyl-d-aspartate (NMDA) receptor by abolishing the Mg^{2+} block in the ion channel which would prolong the opening time of the channel, thus increasing intracellular Ca^{2+} concentration in the neurons (Sen et al., 2007). The characteristics of gatifloxacin transport across blood brain barrier were investigated using primary cultured rat brain microvessel endothelial cells (rBMECs) as an *in vitro* model and study suggested that gatifloxacin transport across rBMECs involves a

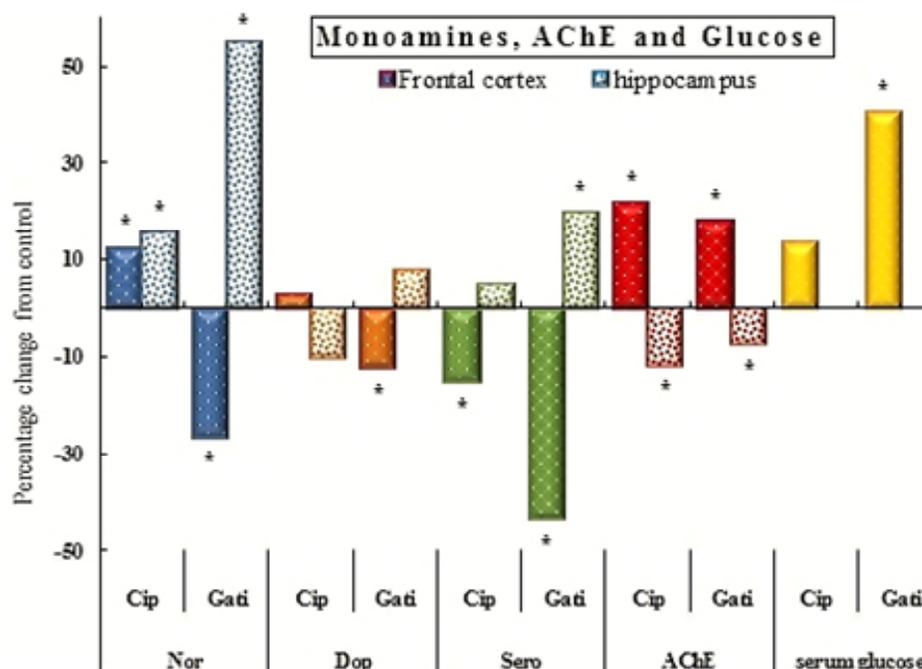


Figure 3. Percentage change from control of monoamines and acetylcholinesterase (AChE) in cortices and hippocampi and serum glucose of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

Na⁺/Ca²⁺ exchange mechanism and extracellular Ca²⁺ (Li et al., 2009). The effect on Ca²⁺ may declare the effect of both antibiotics on taurine levels detected favoring recovery after neuronal hyperactivity (Rawi et al., 2011). Elevated aspartate, serine and glycine might suggest to the excitatory potencies of fluoroquinolones through their activation role on N-Methyl-D-aspartate-type glutamate receptor (NMDAR) (Curras and Dingledine, 1992; Wolosker, 2006; Wolosker et al., 2008).

The regional differences in GABA levels and acetylcholinesterase activities recorded decrease of GABA level and increase of AChE activity in the cortical area. Meanwhile, increase of GABA level and decrease of AChE activity in the hippocampal area in both treatments mimics that predicted in rat epileptic models (Appleyard et al., 1986) and support the proconvulsant effect of the quinolones previously discussed (Smolders et al., 2002; Abdel-Rahman et al., 2013; Arafa et al., 2013). Biochemical studies proposed role for AChE in brain mechanisms in development of status epilepticus through decrease in the AChE activity in the hippocampus (Freitas et al., 2006). The effect of ciprofloxacin and gatifloxacin on GABA levels and acetylcholinesterase activities in cortex and hippocampus and their relation to anxiety and seizure generation was discussed in our previous study (Rawi et al., 2011; Abdel-Rahman et al., 2013). Seizure induction or decrease seizure threshold

related effect to either ciprofloxacin or gatifloxacin single dose was previously declared (Darwish, 2008; Quigley and Lederman, 2004). In addition, serine elevation might be related to hippocampal serotonin increment detected in our study (Santini et al., 2014). Histidine content decreased in the frontal cortex and increased in hippocampus of ciprofloxacin and gatifloxacin treated groups. Histamine synthesis rate is a function of histidine content and histidine raises the possibility of a profound direct effect on CNS function (Yoshimatsu et al., 2002) and the herein results support the anaphylactoid reactions and hypotensive action of quinolones under therapeutic conditions as reported by Furuhashi et al. (1998), Johannes et al. (2007) and Jones et al. (2013).

Fluoroquinolone-associated anaphylaxis may occur after first-ever intake of the agent (Sachs et al., 2006). In addition, drugs that release histamine may provoke headache, asthma, hypotension, arrhythmia, urticaria, pruritus, flushing and other conditions in patients with histamine intolerance (Maintz and Novak, 2007). Monoamines levels recorded in the tested antibiotics shows elevation in noradrenaline and reduction in dopamine and serotonin in the frontal cortex in the Cip and Gati groups except reduced level of noradrenalin in Gati group. However, in hippocampus there are elevations in monoamines levels in both groups except reduction of dopamine level in Gati group. These data

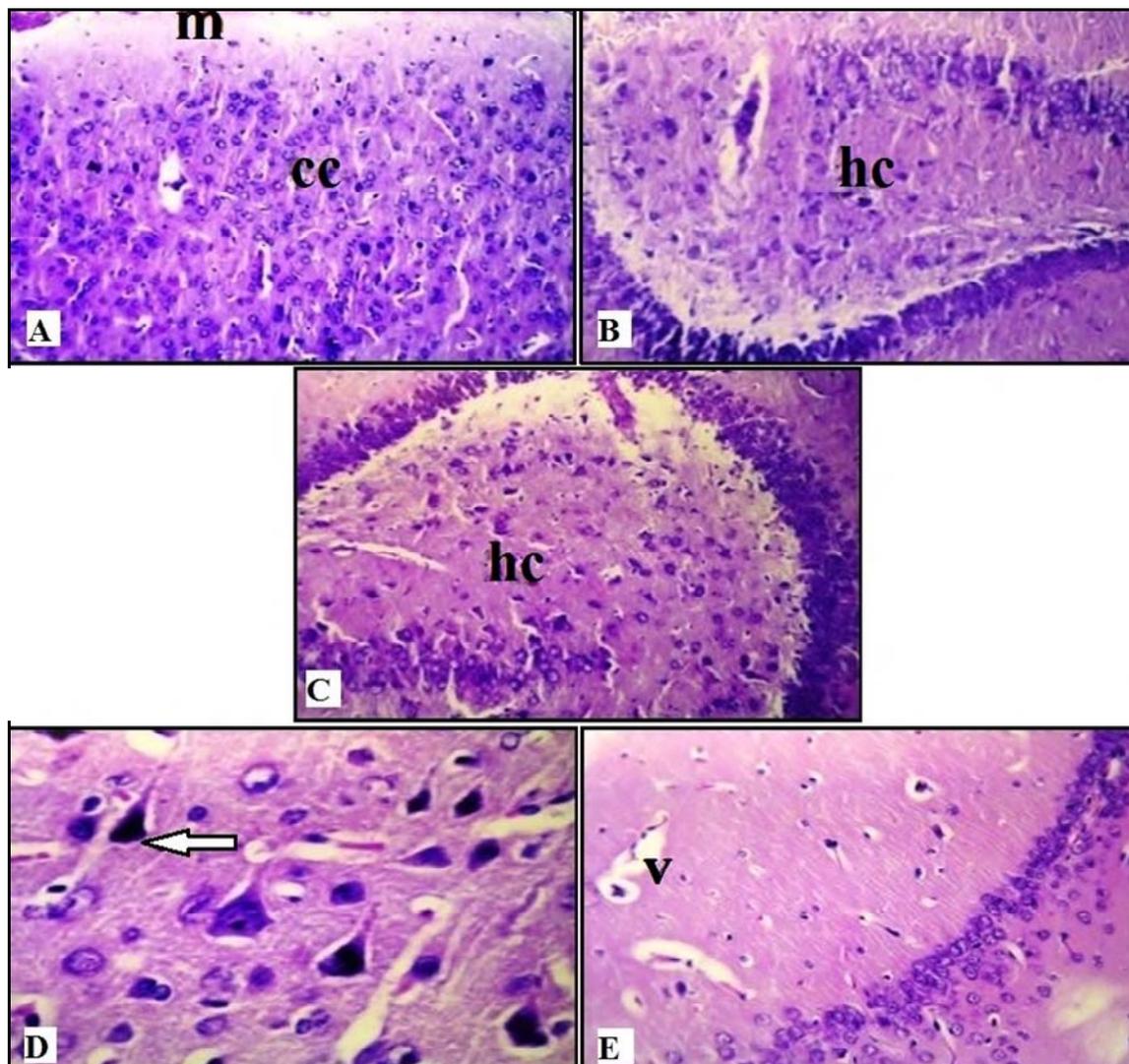


Figure 4. Light micrographs of brain sections in treated groups showing in control group normal histology of cerebral cortex (CC) and covering meninges (m) (H&E \times 40) (A) and normal rat histology of hippocampus (hc). (H&E \times 40) (B). Normal histological structure observed in the hippocampus tissue (hc) in ciprofloxacin group. (H&E \times 40) (C). Neuronal degeneration in the cerebral cortex (arrow). (H&E \times 160) (D), and congestion in the blood vessels and capillaries of the hippocampus (H&E \times 64) (E) in gatifloxacin group.

may be validated by the seizure induction through the assumption about the pharmacological treatments that lowering monoamine levels in the brain generally increase the susceptibility to seizures, while treatments that increase monoamines decrease the susceptibility (Kiyofumi and Akitane, 1977). The data recorded about monoamines in the tested antibiotics may be a supplement data to the previously mentioned seizure inducing activity of quinolones (Ooie et al., 1997; Moorthy et al., 2008; Agbaht et al., 2009). The involvement of prefrontal cortex in depression and the link between

reduced serotonin level in prefrontal cortex and depression symptoms as previously stated (Juckel et al., 1999; Koenigs and Grafman, 2009) is in accordance with the levels detected in our study. So the increment in the intracellular Ca^{2+} ions led to the rupture of the vesicles in the presynaptic terminals and increased the release of the neurotransmitters (Bullock et al., 1995) as a result, the content of catecholamine is decreased. The neurotransmitters alterations support the hyperexcitability which reflected on the histopathology of cortex and hippocampus mainly in the most affected Gati group in

line with several previous studies discussed in Rawi et al. (2011) and Arafa et al. (2013).

As regard to the effect on glucose level in tested groups, Yamada et al. (2006) reported the effect of gatifloxacin on insulin secretion and islet insulin content by using isolated mouse pancreatic islets. Islet insulin content significantly decreased by gatifloxacin already at day one; however, there are some case reports that show that only one or two doses of gatifloxacin can induce hyperglycemia (Biggs, 2003; Arce et al., 2004). Gatifloxacin was withdrawn from clinical use after reports of drug-induced hyperglycemia and other fluoroquinolones reported to interfere with glucose homeostasis (Telfer, 2014). Onyenwenyi et al. (2008) indicated that non-diabetic gatifloxacin treated patients appeared to have an increased risk of hyperglycemia and the risk reduced in diabetics. Ghaly et al. (2009) previously documented that fluoroquinolones did not stimulate insulin secretion in the presence of a basal glucose concentration; rather, they only enhanced the secretion elicited by a stimulatory glucose concentration.

Recent study by Ghaly et al. (2014) explained why fluoroquinolones produce hypo- and hyperglycaemias, because fluoroquinolones affect the function of the mitochondria in pancreatic beta cells, which may diminish the insulinotropic effect of KATP channel closure and contribute to the hyperglycaemic episodes. In addition, ciprofloxacin and gatifloxacin cause oxidative stress and decrease the mitochondrial membrane potential (Loves et al., 2009; Talla and Veerareddy, 2011; Rawi et al., 2011; Arafa et al., 2013). Gatifloxacin acutely diminish gluconeogenesis by inhibition of mitochondrial pyruvate transport (Drozak et al., 2008) since pancreatic beta cells have an exceedingly low antioxidant capacity (Lenzen et al., 1996) and inhibition of pyruvate transport may interfere with nutrient stimulation of insulin secretion. The study of Telfer (2014) extended to suggest a connection between the ingestion of fluoroquinolones antibiotics and the development of type 2 diabetes and advice that follow-up longitudinal studies to be undertaken to examine the history of individual diabetic patients for previous fluoroquinolone exposure. Glucose resuscitation resulting in hyperglycaemia activates the NADPH pathway in neurons, causing cytotoxic oxidative stress. The same phenomenon could also adversely affect oligodendrocytes (Suh et al., 2007). The study concluded from the recorded results that the excitatory potency of ciprofloxacin and gatifloxacin could be achieved from the first dose.

ACKNOWLEDGEMENT

The authors gratefully acknowledge the Department of physiology, NODCAR, Egypt.

Conflict of interests

The authors have declared that no competing interest exists.

REFERENCES

- Abdel-Rahman M, Arafa NM, El-Khadragy MF, Kassab RB (2013). The neuroprotective role of *Nigella sativa* extract on ciprofloxacin and pentylentetrazole treated rats. *Afr. J. Pharm. Pharmacol.* 7(24):1660-1670.
- Agbaht K, Bitik B, Piskinpasa S, Bayraktar M, Topeli A (2009). Ciprofloxacin-associated seizures in a patient with underlying thyrotoxicosis: case report and literature review. *Int. J. Clin. Pharmacol. Ther.* 47(5):303-310.
- Akahane K, Kato M, Takayama S (1993). Involvement of Inhibitory and Excitatory Neurotransmitters in Levofloxacin- and Ciprofloxacin-Induced Convulsions in Mice. *Antimicrob. Agents Chemother.* 37(9):1764-1770.
- Alborzi A, Oskoe S, Pourabbas B, Alborzi S, Astaneh B, Gooya MM, Kaviani MJ (2008). Meningococcal carrier rate before and after hajj pilgrimage: effect of single dose ciprofloxacin on carriage. *East Mediterr. Health J.* 14(2):277-282.
- Ang CW, Carlson GC, Coulter DA (2006). Massive and specific dysregulation of direct cortical input to the hippocampus in temporal lobe epilepsy. *J. Neurosci.* 26(46):11850-11856.
- Appleyard ME, Green AR, Smith AD (1986). Acetylcholinesterase activity in regions of the rat brain following a convulsion. *J. Neurochem.* 46:1789-1793.
- Arafa NM, Abdel-Rahman M, El-Khadragy MF, Kassab RB (2013). Evaluation of the Possible Epileptogenic Activity of Ciprofloxacin: The Role of *Nigella sativa* on Amino Acids Neurotransmitters. *Neurochem. Res.* 8:174-185.
- Arce FCA, Bhasin RS, Pasmantier RM (2004). Severe hyperglycemia during gatifloxacin therapy in patients without diabetes. *Endocr. Pract.* 10:40-44.
- Banchroft JD, Stevens A, Turner DR (1996). Theory and practice of histological techniques. Fourth Ed. Churchill Livingstone, New York, London, San Francisco, Tokyo. Elsevier Health Sciences p 125.
- Biggs WS (2003). Hypoglycemia and hyperglycemia associated with gatifloxacin use in elderly patients. *J. Am. Board Fam. Pract.* 16:455-457.
- Boy D, Well M, Kinzig-Schippers M, Sörgel F, Ankel-Fuchs D, Naber KG (2004). Urinary bactericidal activity, urinary excretion and plasma concentrations of gatifloxacin (400 mg) versus ciprofloxacin (500 mg) in healthy volunteers after a single oral dose. *Int. J. Antimicrob. Agents* 23(1):6-16.
- Bullock J, Boyle J, Wang MB (1995). Synaptic transmission. In: *Physiology Middle East Edition* 3rd ed, Chapter 3, Williams & Wilkins. London pp. 22-31.
- Cavalheiro EA, Fernandes MJ, Turski L, Naffah-Mazzacoratti MG (2006). Spontaneous recurrent seizures in rats: Amino acid and monoamine determination in the hippocampus. *Epilepsia.* 35(1):1-11.
- Curras MC, Dingle R (1992). Selectivity of amino acid transmitters acting at N-methyl-D-aspartate and amino-3-hydroxy-5-methyl-4-isoxazolepropionate receptors. *Mol. Pharmacol.* 41:520-526.
- Darwish T (2008). Ciprofloxacin-induced seizures in a healthy patient. *N. Z. Med. J.* 121(1277):104-105.
- De Sarro A, De sarro G (2001). Adverse reactions to fluoroquinolones. An overview on mechanistic aspects. *Cur. Med. Chem.* 8(4):371-384.
- De Sarro A, Cecchetti V, Fravolini V, Naccari F, Tabarrini O, De sarro G (1999). Effects of novel 6-desfluoroquinolones and classic quinolones on pentylentetrazole-induced seizures in mice. *Antimicrob. Agents. Chemother.* 43(7):1729-1736.
- Drozak J, Miecznik A, Jarzyna R, Bryla J (2008). The inhibition of gluconeogenesis by gatifloxacin may contribute to its hypoglycaemic action

- Eur. J. Pharmacol. 594:39-43.
- Ellman GL, Courtney KD, Andres V, Featherstone RM (1961). A new and rapid colorimetric determination of cholinesterase activity. *Biochem. Pharmacol.* 7:88-95.
- Freitas RM, Sousa FC, Viana GS, Fonteles MM (2006). Acetylcholinesterase activities in hippocampus, frontal cortex and striatum of Wistar rats after pilocarpine-induced status epilepticus. *Neurosci. Lett.* 399 (1-2):76-78. PMID: 16481111
- Freitas RM, Vasconcelos SMM, Souza FCF, Viana GSB, Fonteles MMF (2004). Monoamine levels after Pilocarpine-induced status epilepticus in hippocampus and frontal cortex of wistar rats. *Neurosci. Lett.* 370 (2-3):196-200.
- Furuhata K, Hayakawa H, Soumi K, Arai H, Watanabe Y, Narita H (1998). Histamine-releasing properties of T-3762, a novel fluoroquinolone antimicrobial agent in intravenous use. I. Effects of doses and infusion rate on blood pressure, heart rate and plasma histamine concentration. *Biol. Pharm. Bull.* 21(5):456-460.
- Ghaly H, Jörns A, Rustenbeck I (2014). Effect of fluoroquinolones on mitochondrial function in pancreatic beta cells. *Eur. J. Pharm. Sci.* 52:206-214.
- Ghaly H, Kriete C, Sahin S, Pflöger A, Holzgrabe U, Zünkler BJ, Rustenbeck I (2009). The insulinotropic effect of fluoroquinolones. *Biochem. Pharmacol.* 77:1040-1052.
- Gorun V, Proinov I, Baltescu V, Balaban G, Barzu O (1978). Modified Ellman procedure for assay of cholinesterase in crude enzymatic preparation. *Anal. Biochem.* 86:324-326.
- Guidance for Industry and Reviewers (2002). Estimating the safe starting dose in clinical trials for therapeutics in adult healthy volunteers. Food and Drug Administration 1-26. <http://www.fda.gov/OHRMS/DOCKETS/98fr/02d-0492-gdI0001-vol1.pdf>
- Heidari Bateni Z, Shahrokh H, Salimi H, Safari H, Tabatabai M, Saedi D (2014). Single-Dose versus Multiple-Dose Ciprofloxacin plus Metronidazole Prophylaxis in Transrectal Ultrasound-Guided Biopsy of the Prostate: a Randomized Controlled Trial. *Acta Med. Iran.* 52(9):664-670.
- Heinrikson RL, Meredith SC (1984). Amino acid analysis by RP-HPLC: precolumn Derivatization with phenylisothiocyanate. *Anal. Biochem.* 136:65-74.
- Johannes CB, Ziyadeh N, Seeger JD, Tucker E, Reiter C, Faich G (2007). Incidence of Allergic Reactions Associated with Antibacterial Use in a Large, Managed Care Organisation. *Drug Saf.* 30(8):705-713.
- Jones SC, Budnitz DS, Sorbello A, Mehta H (2013). US-based emergency department visits for fluoroquinolone-associated hypersensitivity reactions. *Pharmacoepidemiol. Drug Saf.* 22(10):1099-1106.
- Jose J, Jimmy B, Saravu K (2007). Dysglycemia associated with the use of fluoroquinolones-focus on gatifloxacin. *J. Clin. Diagn. Res.* (3):185-187.
- Juckel G, Mendlin A, Jacobs BL (1999). Electrical stimulation of rat medial prefrontal cortex enhances forebrain serotonin output: implications for electroconvulsive therapy and transcranial magnetic stimulation in depression. *Neuropsychopharmacology* 21(3):391-398.
- Kaushik JS, Gupta P, Faridi MM, Das S (2010). Single dose azithromycin versus ciprofloxacin for cholera in children: a randomized controlled trial. *Indian Pediatr.* 47(4):309-315.
- Kelly ME, Batty RA, McIntyre DC (1999). Cortical spreading depression reversibly disrupts convulsive motor seizure expression in amygdala-kindled rats. *Neuroscience* 91:305-313.
- Kelly ME, Staines WA, McIntyre DC (2002). Secondary generalization of hippocampal kindled seizures in rats: examination of the periform cortex. *Brain Res.* 957(1):152-161.
- Kiyofumi K, Akitane M (1977). Brain Monoamines in Seizure Mechanism (Review). *Folia Psychiatr. Neurol. Jpn.* 31(3):483-489.
- Koenigs M, Grafman J (2009). The functional neuroanatomy of depression: distinct roles for ventromedial and dorsolateral prefrontal cortex. *Behav. Brain Res.* 201(2):239-243.
- Lenzen S, Drinkgern J, Tiedge M (1996). Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. *Free Radic Biol. Med.* 20(3):463-466.
- Li Y, Liu L, Li J, Xie L, Wang GJ, Liu XD (2009). Transport of gatifloxacin involves Na⁺/Ca²⁺ exchange and excludes P-glycoprotein and multidrug resistance associated-proteins in primary cultured rat brain endothelial cells. *Eur. J. Pharmacol.* 616(1-3):68-72
- Loo PS, Ridgway GL, Oriol JD (1985). Single dose ciprofloxacin for treating gonococcal infections in men. *Genitourin Med.* 61(5):302-305.
- Lowes DA, Wallace C, Murphy MP, Webster NR, Galley HF (2009). The mitochondria targeted antioxidant MitoQ protects against fluoroquinolone-induced oxidative stress and mitochondrial membrane damage in human Achilles tendon cells. *Free Radic Res.* 43(4): 323-328.
- Maintz L, Novak N. (2007). Histamine and histamine intolerance. *Am. J. Clin. Nutr.* 85(5):1185-1196.
- Moorthy N, Raghavendra N, Venkatarathnamma PN (2008). Levofloxacin-induced acute psychosis. *Indian J. Psychiatry* 50(1):57-58.
- Motomura M, Kataoka Y, Takeo G, Shibayama K, Ohishi K, Nakamura T, Niwa M, Tsujihata M, Nagataki S (1991). Hippocampus and frontal cortex are the potential mediatory sites for convulsions induced by new quinolones and non-steroidal anti-inflammatory drugs. *Int. J. Clin. Pharmacol. Ther. Toxicol.* 29(6):223-227.
- Naber KG, Allin DM, Clarysse L, Haworth DA, James IG, Raini C, Schneider H, Wall A, Weitz P, Hopkins G, Ankel-Fuchs D (2004). Gatifloxacin 400 mg as a single shot or 200 mg once daily for 3 days is as effective as ciprofloxacin 250 mg twice daily for the treatment of patients with uncomplicated urinary tract infections. *Int. J. Antimicrob. Agents* 23(6):596-605.
- Onyenwenyi AJ, Winterstein AG, Hatton RC (2008). An evaluation of the effects of gatifloxacin on glucose homeostasis. *Pharm. World Sci.* 30(5):544-549.
- Ooie T, Terasaki T, Suzuki H, Sugiyama Y (1997). Quantitative brain microdialysis study on the mechanism of quinolones distribution in the central nervous system. *Drug Metab. Dispos.* 25(7):784-789.
- Pagel P, Blome J, Wolf HU (2000). High-performance liquid chromatographic separation and measurement of various biogenic compounds possibly involved in the pathomechanism of Parkinson's disease. *J. Chromatogr. B Biomed. Sci. Appl.* 746(2): 297-304.
- Quigley CA, Lederman JR (2004). Possible gatifloxacin-induced seizure. *Ann. Pharmacother.* 38(2):235-237.
- Rawi SM, Arafa NMS, El-Hazmi MM (2011). Evaluation of the Effects of Ciprofloxacin or Gatifloxacin on Neurotransmitters Levels in Rat Cortex and Hippocampus. *Afr. J. Pharm. Pharmacol.* 5:993-1005.
- Sachs B, Riegel S, Seebeck J, Beier R, Schichler D, Barger A, Merk HF, Erdmann S (2006). Fluoroquinolone-associated anaphylaxis in spontaneous adverse drug reaction reports in Germany: differences in reporting rates between individual fluoroquinolones and occurrence after first-ever use. *Drug Saf.* 29(11):1087-1100.
- Santini MA, Balu DT, Puhl MD, Hill-Smith TE, Berg AR, Lucki I, Mikkelsen JD, Coyle JT (2014). D-serine deficiency attenuates the behavioral and cellular effects induced by the hallucinogenic 5-HT_{2A} receptor agonist DOI. *Behav. Brain Res.* 259:242-246.
- Schmuck G, Schürmann A, Schlüter G. (1998). Determination of the excitatory potencies of fluoroquinolones in the central nervous system by an *in vitro* model. *Antimicrob. Agents Chemother.* 42(7):1831-1836.
- Sen S, Jaiswal AK, Yanpallewar S, Acharya SB (2007). Anxiogenic potential of ciprofloxacin and norfloxacin in rats. *Singapore Med. J.* 48(11):1028-1032.
- Smolders I, Gousseau C, Marchand S, Couet W, Ebinger G, Michotte Y (2002). Convulsant and Subconvulsant Doses of Norfloxacin in the Presence and Absence of Biphenylacetic Acid Alter Extracellular Hippocampal Glutamate but Not Gamma-Aminobutyric Acid Levels in Conscious Rats. *Antimicrob. Agents Chemother.* 46(2):471-477.
- Suh W S, Gun ET, Hamby AM, Chan PH, Swanson RA (2007). Hypo-

- glycemic neuronal death is triggered by glucose reperfusion and activation of neuronal NADPH oxidase. *J. Clin. Invest.* 117(4):910–918.
- Talla V, Veerareddy P (2011). Oxidative stress induced by fluoroquinolones on treatment for complicated urinary tract infections in Indian patients. *J. Young Pharm.* 3(4):304-309.
- Telfer SJ (2014) Fluoroquinolone antibiotics and type 2 diabetes mellitus. *Med. Hypotheses.* 83(3):263-269.
- Terzi C, Kiliç D, Unek T, Hoşgörler F, Füzün M, Ergör G (2005). Single-dose oral ciprofloxacin compared with single-dose intravenous cefazolin for prophylaxis in inguinal hernia repair: a controlled randomized clinical study. *J. Hosp. Infect.* 60(4):340-347.
- Wolosker H, Dumin E, Balan L, Foltyn VN (2008). D-amino acids in the brain: D-serine in neurotransmission and neurodegeneration. *FEBS J.* 275(14):3514-3526.
- Wolosker H (2006). D-serine regulation of NMDA receptor activity. *Sci STKE* (356):pe41.
- Yamada C, Nagashima K, Takahashi A, Ueno H, Kawasaki Y, Yamada Y, Seino Y, Inagaki N (2006). Gatifloxacin acutely stimulates insulin secretion and chronically suppresses insulin biosynthesis. *Eur. J. Pharmacol.* 553(1-3):67-72.
- Yoshimatsu H, Chiba S, Tajima D, Akeh Y, Sakata T (2002). Histidine suppresses food intake through its conversion into neuronal histamine. *Exp. Biol. Med. (Maywood).* 227(1):63-68.