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Full Length Research Paper

Meta-analysis of methylenetetrahydrofolate reductase (MTHFR) A1298C polymorphism and risk of orofacial cleft

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Polymorphisms in key genes involving the folate pathway have been reported to be associated with the risk of orofacial cleft (OFC) and several studies were published with conflicting results. A meta-analysis of the previous studies of allelic association between OFC with A1298C polymorphisms of the methylenetetrahydrofolate reductase (MTHFR) gene was carried out. Odds ratios (ORs) with 95% confidence intervals (Cls) were estimated to assess the association between MTHFR A1298C polymorphism and OFC risk. A total of 11 studies including 1628 cases and 2676 controls were involved in this meta-analysis. No statistical relationship was found with any genetic model (C vs. A (Additive): OR = 1.14, 95%Cl = 0.76-1.65, P = 0.47; CC vs. AA (homozygote): OR = 0.90, 95%Cl = 0.72-1.15, P = 0.0.41; AC vs. AA (co-dominant): OR = 0.97, 95%Cl = 0.85-1.11, P = 0.0.63; CC+AC vs. AA (Dominant): OR = 0.96, 95%Cl = 0.84-1.1, P = 0.51; CC vs. AC+AA (Recessive): OR = 0.93, 95%Cl = 0.74-1.16, P = 0.52). The present meta-analysis supports that the common A1298C polymorphism of MTHFR gene is not risk factor for OFC.

Key words: Orofacial cleft, cleft lip, cleft palate, methylenetetrahydrofolate reductase (MTHFR), A1298C, folic acid.

INTRODUCTION

Approximately 90% of craniofacial congenital abnormalities comprised orofacial cleft (OFC) or cleft lip and/or palate (CL/P). According to the world health organization (WHO) data, the frequency of this pathology in the world is 0.6 to 1.6 cases per 1000 newborns (Shaw et al., 2001; Chorna et al., 2011). Prevalence rate varies according to geographical origin, sex, racial background, ethnicity, and socio-economic status (Vanderas et al.,

1987; Croen et al., 1998; Clark et al., 2003; Brito et al., 2011; Aslar et al., 2013). Prenatal folic acid supplementation to pregnant women has been shown to reduce the incidence of CL in many (van Rooij et al., 2004; Badovinac et al., 2007; Rouget et al., 2005; Yazdy et al., 2007; Wilcox et al., 2007), but not all (Ray et al., 2003) populations studied (Sozen et al., 2009). Several studies established that polymorphisms in genes implicated

*Corresponding author. E-mail: raivandana@rediffmail.com. Fax: 05452-252244 or 05452-252344. Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> License 4.0 International License in folate metabolism may play a significant role in the OFC etiology. Among several genes that take part in folate metabolism, the methylenetetrahydrofolate reductase gene (MTHFR) has been the most frequently reported to be associated with OFC.

MTHFR (EC.1.5.1.20) is a key enzyme in folate and homocysteine metabolism and catalyzes the reduction of 10-methylenetetrahydrofolate 5. to methyltetrahydrofolate, which provides the methyl group for the remethylation of homocysteine to methionine. Methionine is in turn converted to S-adenosylmethionine (SAM), the common methyl donor for the methylation processes of DNA, proteins, phospholipids and neurotransmitters (Finkelstein, 1990; Bailey and Gregory, 1999; Ozarda et al., 2009). The MTHFR gene is localized on chromosome 1p36.3 and two common and clinically important polymorphisms (C677T and A1298C) identified in the MTHFR gene (Frosst et al., 1995; Weisberg et al., 1998) are implicated in the development of OFC. MTHFR C677T polymorphism is very well studied but A1298C polymorphism is less explored. A1298C influences specific activity of the enzyme, homocysteine levels, and plasma folate concentration, but to a lesser extent than the C677T polymorphism does (Blount et al., 1997; Shen et al., 2005).

Substitution at nucleotide 1298 (exon 7) results in an amino acid substitution of glutamate for alanine at codon 429. A1298C (glutamate to alanine) polymorphism, has been associated with decreased enzyme activity (40%), although to a lesser extent than C677T (Weisberg et al., 1998). Those who have the AC or CC genotype present with a decreased ability to produce the methyl form of folate, which together with cobalamin, is essential for the remethylation of homocysteine to methionine. The resulting abnormality in folate metabolism and the resultant increase in homocysteine levels may be a direct cause of the observed teratogenicity, homocysteine itself may be toxic to the embryo or it may be an indicator of reduced availability of SAM for the methylation of DNA. Animal studies suggest that a decreased conversion of homocysteine to methionine could be a crucial step in causing neural tube defects. It has been shown that rat embryos in culture require methionine for neural tube closure (Mills et al., 1999).

A1298C allele frequency differs greatly in various ethnic groups of the world. The prevalence of the A1298C homozygote variant genotype ranges from 7 to 12% in White populations from North America and Europe. Lower frequencies have been reported in Hispanics (4 to 5%), Chinese (1 to 4%) and Asian populations (1 to 4%) (Botto and Yang, 2000; Rabein and Ulrich, 2003). A number of molecular epidemiological studies have been conducted to investigate the associations of the MTHFR A1298C polymorphism with OFC. However, the results remain conflicting rather than conclusive. Hence, a meta-analysis to derive a more precise estimation of this association is needed. In light of the above facts, a meta-analysis of all available studies relating the A1298C polymorphism of MTHFR gene to the risk of having cleft lip was conducted.

METHODOLOGY

Selection of studies

All studies that investigate the association of the A1298C polymorphism in the MTHFR gene with CLP published before October 2013 were considered in the meta-analysis. The studies were identified by extended computer-based searches of the PubMed, Google Scholar, Elsevier and Springer Link databases. As a search criterion, the following terms were used: 'MTHFR', 'orofacial cleft', 'OFC', 'cleft lip', 'cleft lip and palate', 'A1298C'.

The following inclusion criteria were used: (i) studies must have a case–control study, (ii) study must be published as full papers, (iii) authors must investigate patients with cleft lip and palate cases and healthy control subjects, (iv) authors must provide information on genotype frequencies of the MTHFR A1298C polymorphism or sufficient data for the calculation, (iv) studies with overlapping cases and/or controls, the largest study with extractable data was included. The major reasons for exclusion of studies were (1) only case studied, (2) review papers, editorial, letter to editor and (3) containing overlapping data.

Data extraction

Following information was extracted from each study: first author, journal, year of publication, racial descent of study population, demographics, matching, validity of the genotyping method, and the number of cases and controls for MTHFR A1298C. The frequencies of A and C alleles were calculated for cases and controls from the corresponding genotype distributions.

Meta-analysis

The meta-analysis examined the overall association of the C allele with the risk of OFC relative to the A allele, the additive model for C allele (C vs. A), the co-dominance model (AC vs. AA), the homozygote model for allele C (CC vs. AA), the dominant model for C allele (CC + AC vs. AA), and the recessive model for C allele (CC vs. AC + AA). All associations were indicated as odds ratios (OR) with the corresponding 95% confidence interval (CI). A pooled OR was estimated based on the individual ORs. Heterogeneity between studies was tested using the Q statistic (Cochran, 1954). Heterogeneity was considered statistically significant if P<0.05. Heterogeneity was quantified with the I^2 metric, which is independent of the number of studies in the meta-analysis (I²<25%: no heterogeneity; I²=25-50%: moderate heterogeneity; I²=50-75%: large heterogeneity; 12>75%: extreme heterogeneity (Higgins and Thompson, 2002). The pooled OR was estimated using fixed effects (Mantel and Haenszel, 1959) and random effects (Dersimonian and Laird, 1986) models (Whitehead, 2002). Random effects modeling assume a genuine diversity in the results of the studies, and it incorporates the calculations of between-study variability; it therefore tends to provide wider CIs (Zintzaras and Hadjigeorgiou, 2005).

Publication bias

An estimate of the potential publication bias was carried out by

Table 1.	Characteristics	of eleven	studies	included	in the	present	meta-analy	/sis.
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Study	Country	Year	Case	Control	Reference
Tolarova et al.	Argentina	1998	108	103	Am J Hum Genet, 63:A27.
Grunert et al.,	Germany	2002	65	184	Mund Kiefer Gesichtschir 6:131–133.
Shoteresuk et al.	Thailand	2003	109	202	J Med Genet, 40:e64.
van Roij et al.	The Netherlands	2003	94	115	Am J Epidemiol, 157:583–591.
Pezzetti et al.	Italy	2004	110	289	Hum Mutat, 24:104–105.
Mills et al.	Ireland	2008	407	1050	Birth Defects Research, 82:636–643.
Ali et al.	India	2009	323	214	Genetic Testing and Molecular Biomarkers, 13(3).
Sozen et al.	Venezuela	2009	179	138	J. Genet. Genomics, 36: 283-288.
Chorna et al.	Ukraine	2011	33	50	Cytology and Genetics, 45: 177–181
Semic-Jusufagic et al.	Turkey	2012	56	76	The Turkish Journal of Pediatrics, 54: 617-625
Kumari et al.	India	2013	467	469	J. Biosci., 38: 21–26.

funnel plot, in which the standard error (SE) of log (OR) of each study was plotted against its log (OR). An asymmetric plot suggested a possible publication bias. The funnel plot asymmetry was assessed by Egger's test, and P=0.05 was considered representative of statistically significant publication bias (Egger et al., 1997). All analyses were performed using the computer program MIX version 1.7 (Bax et al., 2006). A p value less than 0.05 was considered statistically significant, and all the p values were two sided.

RESULTS

Eligible studies

Eleven articles were found to be eligible for the inclusion in the present meta-analysis (Tolarova et al., 1998; Grunert et al., 2002; Shoteresuk et al., 2003; van Roij et al., 2003; Pezzetti et al., 2004; Mills et al., 2008; Ali et al., 2009; Sozen et al., 2009; Chorna et al., 2011; Semic-Jusufagic et al., 2012; Kumari et al., 2013). All these eleven studies were performed in different countries: Argentina (Tolarova et al., 1998), Germany (Grunert et al., 2002), India (Ali et al., 2009; Kumari et al., 2013), Ireland (Mills et al., 2008), Italy (Pezzetti et al., 2004), Netherlands (van Roij et al., 2003), Thailand (Shoteresuk et al., 2003), Ukraine (Chorna et al., 2011), and Venezuela (Sozen et al., 2009). In one study, Ali et al. (2009) reported only allele numbers (Table 1).

Statistical analysis

Overall, eleven studies provided 1628/2676 cases/controls for MTHFR A1298C. The frequencies of the genotypes MTHFR 1298AA and AC were the highest in both OFC cases and controls, and allele A was the most common. In all eleven studies, total cases were 1628 with AA (825), AC (668) and CC (135), and controls were 2676 with AA (1285), AC (1140), and CC (251) genotypes. In controls genotypes, percentage of AA, AC

and CC were 48, 42.6, and 9.38%, respectively. In total cases, genotype percentage of AA, AC, and CC was 50.6, 41 and 8.3%, respectively. The genotype and allele distributions are as shown in Table 2. Only in four studies (Shoteresuk et al., 2003; van Roij et al., 2003; Semic-Jusufagic et al., 2012; Kumari et al., 2013), OR was above one and in other seven studies did not show any association between MTHFR A1298C polymorphism and OFC. The distribution of genotypes in the control groups were in Hardy-Weinberg equilibrium (HWE) in all studies. Lack of HWE indicates possible genotyping errors and/or population stratification (Zintzaras, 2007).

Allele contrast meta-analysis

The main results of this meta-analysis and the heterogeneity test were shown in Tables 3. Mutant allele (C vs. A) did not show significant association with OFC in both fixed effect (OR=1.06, 95% CI=0.96-1.16, P=0.25, P_{hetero} <0.0001, I²=92.62%, P_{Pb} =0.60) and random effect (OR=1.14, 95% CI=0.79-1.65, P=0.47) models (Table 3 and Figure 1).

Genotype contrast meta-analysis

Overall, no significantly elevated cleft lip risk was detected in any genetic models when all studies were pooled into the meta-analysis. Homozygote model (CC vs. AA) did not show significant association with OFC in both fixed effect (OR=0.97, 95% CI=0.72-1.15, P=0.41, P_{hetero} =0.04, I²=47.04%, P_{Pb} =0.57) and random effect (OR=0.88, 95% CI=0.59-1.28, P=0.49) models (Table 3 and Figure 2). Similarly dominant model (CC+AC vs. AA) also did not show any association between A1298C polymorphism and risk of OFC either with fixed effect (OR=0.96, 95% CI=0.84-1.08, P=0.51, P_{hetero}=0.13, I²=34.65%, P_{Pb}=0.61) or random effect (OR=0.94, 95% CI=0.78-1.12, P=0.47) model. Meta-analysis result using

	Genotype					Alleles				
Study ID	AA		AC		CC		Α		С	
	Case	Control	Case	Control	Case	Control	Case	Control	Case	Control
Tolarova et al. (1998)	67	63	39	33	2	7	173	43	159	47
Grunert et al. (2002)	28	77	30	80	7	27	86	44	234	134
Shoteresuk et al. (2003)	55	108	48	80	6	14	158	60	296	108
van Roij et al. (2003)	48	61	34	43	12	11	130	58	165	65
Pezzetti et al. (2004)	56	95	46	151	8	43	158	62	341	237
Mills et al. (2008)	202	519	172	439	33	92	576	238	1477	623
Ali et al. (2009)	-	-	-	-	-	-	483	295	163	133
Sozen et al. (2009)	138	101	37	33	4	4	313	235	45	41
Chorna et al. (2011)	19	24	12	22	2	4	50	70	16	30
Semic-Jusufagic et al. (2012)	21	36	25	36	10	4	67	108	45	44
Kumari et al. (2013)	191	201	225	223	51	45	607	625	327	313

Table 2. The distributions of MTHFR A1298C genotypes and allele frequencies for CLP cases and controls.

Table 3. Summary estimates for the odds ratio (OR) of MTHFR A1298C in various allele/genotype contrasts, the significance level (p value) of heterogeneity test (Q test), and the I^2 metric: overall analysis, subgroup analyses, and publication bias p-value (Egger test).

Genetic models	Fixed effect OR (95% CI), p	Random effect OR (95% CI), p	Heterogeneity p-value (Q test)	l² (%)	Publication Bias (p of Egger's test)
Allele contrast (C vs. A)	1.06 (0.96-1.16), 0.25	1.14 (79-1.65), 0.47	<0.0001	92.62	0.60
Co-dominant (AC vs. AA)	0.97 (0.85-1.107), 0.63	0.96 (0.83-1.11), 0.61	0.38	6.12	0.56
Homozygote (CC vs. AA)	0.90 (0.72-1.15), 0.41	0.88 (0.59-1.28), 0.49	0.04	47.04	0.57
Dominant (CC+AC vs. AA)	0.96 (0.84-1.08), 0.51	0.94 (0.78-1.12), 0.47	0.13	34.65	0.61
Recessive (AA+AC vs. CC)	0.93 (0.74-1.16), 0.52	0.91 (0.66-1.26), 0.58	0.14	33.64	0.59

co-dominant and recessive genetic models were also not significant: (AC vs. AA: OR=0.96, 95% CI=0.85-1.11, P=0.63, P_{hetero}=0.38, I²=6.16%, P_{Pb}=0.56 and CC vs. AC+AA: OR=0.93, 95% CI=0.74-1.16, P=0.52, P_{hetero}=0.14, I²=33.64%, P_{Pb}=0.59), the pooled ORs were performed using fixed-effect model.

Table 3 lists the main results of the meta-analysis.

Sensitivity analysis

Sensitivity analysis was performed by sequential omission of individual studies from various contrasts.

The associations of the A1298C polymorphism with cleft lip did not change during the sensitivity analysis.

Publication bias

Funnel plots using standard error and precision values for allele and genotypes using fixed effect model were generated (Figure 3). Symmetrical distribution of studies in the funnel plots suggests absence of publication bias. This is also supported by Beggs and Eggers test (Begg's



Figure 1. Forest plots for the association between MTHFR C677T polymorphism and orofacial cleft for additive model (C vs. A) with random effect model



Figure 2. Forest plots for the association between MTHFR C677T polymorphism and orofacial cleft for homozygote model (CC vs. AA) with fixed effect model.

p=0.43, Egger's p=0.60 for C vs. A; Begg's p=0.37, Egger's p=0.57 for CC vs. AA; and Begg's p=0.72, Egger's p=0.56 for AC vs. AA; Begg's p=0.47, Egger's p=0.61 for CC+AC vs. AA; Begg's p=0.85, Egger's p=0.59 for CC vs. AC+AA) (Table 3).

DISCUSSION

Normal MTHFR activity is crucial to maintain the pool of

circulating folate and methionine and to prevent the accumulation of homocysteine (Frosst et al., 1995). Homocysteine considered as a useful and important metabolic marker of the overall folate status. Folic acid derivatives provide essential single carbon units fro nucleic acid synthesis and methylation reactions both of which are essential for cell division, gene expression and maintenance of chromosome structure during fetal development. It is interesting to note that the case control



Figure 3. (A) Funnel plots A. precision versus OR (C vs. A), (B) standard error versus OR (C vs. A), (C) precision versus OR (CC vs. AA), (D) standard error versus OR (CC vs. AA).

studies have indicated an effect of the maternal MTHFR genotype rather than that of the affected child (Martinelli

et al., 2001; Prescott et al., 2002; Pezzetti et al., 2004). The association of MTHFR polymorphisms with the

increased risk of OFC supports the protective effect of maternal use of multivitamins containing folic acid with respect to the occurrence of orofacial clefts (Bailey et al., 2005).

Several meta-analysis studies illustrate the utility of the technique in identifying genes of small effects like MTHFR with phenotypes like-NTD (Zhang et al., 2013); down syndrome (Zintzaras, 2007; Wu et al., 2013); cardiovascular disease (Xuan et al., 2011), stroke (Yadav et al., 2013); migraine (Shurks et al., 2010), Alzheimer's (Zhang et al., 2010), bipolar disorder (Rai, 2011), and depression (Zintzaras, 2006; Wu et al., 2013). Author identified one meta-analysis (Verkleij-Hagoort et al., 2007) published in 2007 concerning similar topic during the literature search. Verkleij-Hagoort performed a metaanalysis based on eight studies and find meager between MTHFR C677T polymorphisms and orofacial cleft (OR=1.01; 95% CI=0.87-1.16; I²=0%). They investigated MTHFR C677T polymorphism and did not investigate A1298C polymorphism.

This study has some limitations and strength also. The main strength was the absence of publication bias and except additive model, low heterogeneity was observed. The insignificant and inconclusive result of the present meta-analysis may be due to (i) small number of studies (only eleven studies), (ii) small sample size, (iii) different ethnic backgrounds of the individuals included in the study, (iv) widely spread exclusion and inclusion criteria which might complicate the comparison between the studies.

In conclusion, result of the present meta-analysis demonstrated that MTHFR A1298C polymorphism did not show any association with CLP and is not a risk factor for oral facial cleft. Oral facial cleft has not been studied as extensively. Further research on facial cleft associations with this MTHFR polymorphism is needed.

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Conflict Interests

The authors declare that there is no conflict of interests regarding the publication of this article.

Abbreviations

OFC, Orofacial cleft; **MTHFR**, methylenetetrahydrofolate reductase C677T ; **SAM S**, adenosylmethionine.

REFERENCES

- Ali A, Singh SK, Raman R (2009). MTHFR 677TT alone and IRF6 820GG together with MTHFR 677CT, but not MTHFR A1298C, are risks for nonsyndromic cleft lip with or without cleft palate in an Indian population. Genet. Test Mol. Biomarkers 13(3):355-60.
- Aslar D, Ozdiler E, Tastan H, Altug AB (2013). Determination of Methylenetetrahydrofolate Reductase (MTHFR) gene polymorphism in Turkish patients with nonsyndromic cleft lip and palate. Int. J. Pediatr. Otorhinolaryngol. 77:1143-1146.
- Badovinac RL, Werler WM, Williams PL, Kelsey KT, Hayes C (2007). Folic acid-containing supplement consumption during pregnancy and risk for oral clefts: A meta-analysis. Clin. Mol. Teratol. 79:8-15.
- Bailey LB, Gregory JF (1999). Folate metabolism and requirements. J. Nutr. 129:779-782.
- Bax L, Yu LM, Ikeda N, Tsuruta H, Moons KG (2006). Development and validation of MIX: comprehensive free software for meta-analysis of causal research data. BMC Med. Res. Methodol. 6:50.
- Botto LD, Yang Q (2000). 5,10-Methylenetetrahydrofolate reductase gene variants and congenital anomalies: a HuGE review. Am. J. Epidemiol. 151(9):862-77.
- Brito LA, Cruz LA, Rocha KM, Barbara LK, Silva CB, Bueno DF, Aguena M, Bertola DR, Franco D, Costa AM, Alonso N, Otto PA, Passos-Bueno MR (2011). Genetic contribution for non-syndromic cleft lip with or without cleft palate (NS CL/P) in different regions of Brazil and implications for association studies. Am. J. Med. Genet. 155A(7):1581-1587.
- Chorna LB, Akopyan HR, Makukh HV, Fedoryk IM (2011). Allelic Polymorphisms in the MTHFR, MTR and MTRR Genes in Patients with Cleft Lip and/or Palate and Their Mothers. Cytol Genet. 45(3):177-181.
- Clark JD, Mossey PA, Sharp L, Little J (2003). Socioeconomic status and orofacial clefts in Scotland, 1989 to 1998. Cleft Palate Craniofac. J. 40:481-485.
- Cochran WG (1954). The combination of estimates from different experiments. Biometrics 10:101-129.
- Croen LA, Shaw GM, Wasserman CR, Tolarova MM (1998). Racial and ethnic variations in the prevalence of orofacial clefts in California 1983-1992. Am. J. Med. Genet. 79:42-47.
- DerSimonian R, Laird N (1986). Meta-analysis in clinical trials. Control Clin. Trials 7:177-88.
- Egger M, Davey Smith G, Schneider M, Minder C (1997). Bias in metaanalysis detected by a simple, graphical test. BMJ 315:629-634.
- Finkelstein JD (1990). Metionine metabolism in mammals. J. Nutr. Biochem. 1:228-237.
- Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, Boers GJH, den Heijer M, Kluijtmans LAJ, van den Heuve LP, Rozen R (1995). A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. Nat. Genet. 10(1):111-113.
- Grunert RR, Braune A, Schnackenberg E, Schloot W, Krause HR (2002). Genetic differences in enzymes of folic acid metabolism in patients with lip-jaw-palate clefts and their relatives. Mund Kiefer Gesichtschir. 6(3):131-133.
- Higgins JP, Thompson SE (2002). Quantifying heterogeneity in a metaanalysis. Stat. Med. 21(11):1539-1558.
- Kumari P, Ali A, Sukla KK, Singh SK, Raman R (2013). Lower incidence of nonsyndromic cleft lip with or without cleft palate in females: Is homocysteine a factor? J. Biosci. 38(1):21-26.
- Mantel N, Haenszel W (1959). Statistical aspects of the analysis of data from retrospective studies of disease. J. Natl. Cancer Inst. 22(4):719-48.
- Martinelli M, Scapoli L, Pezzetti F, Carinci F, Carinci P, Stabellini G, Bisceglia L, Gombos F, Tognon M (2001). C677T variant form at the MTHFR gene and CL/P: A risk factor for mothers? Am. J. Med. Genet. 98(4):357-360.
- Mills JL, Kirke PN, Molloy AM, Burke H, Conley MR, Lee YJ, Mayne PD, Weir DG, Scott JM (1999). Methylenetetrahydrofolate reductase thermolabile variant and oral clefts. Am. J. Med. Genet. 86(1):71-74.
- Mills JL, Molloy AM, Parle-McDermott A, Troendle JF, Brody LC, Conley MR, Cox C, Pangilinan F, Orr DJ, Earley M, McKiernan E, Lynn EC, Doyle A, Scott JM, Kirke PN (2008). Folate-Related Gene

Polymorphisms as Risk Factors for Cleft Lip and Cleft Palate. Clin. Mol. Teratol. 82(9):636-643.

- Ozarda Y, Sucu DK, Hizli B, Aslan D (2009). Rate of T alleles and TT genotype at MTHFR 677C->T locus or C alleles and CC genotype at MTHFR 1298A->C locus among healthy subjects in Turkey: Impact on homocysteine and folic acid status and reference intervals. Cell Biochem. Funct. 27:568-577.
- Pezzetti F, Martinelli M, Scapoli L, Carinci F, Palmieri A, Marchesini J, Carinci P, Caramelli E, Rullo R, Gombos F, Tognon M (2004). Maternal MTHFR Variant Forms Increase the Risk in Offspring of Isolated Nonsyndromic Cleft Lip with Or Without Cleft Palate. Hum. Mutat. 24(1):104-105.
- Prescott NJ, Winter RM, Malcolm S (2002). Maternal MTHFR genotype contributes to the risk of non-syndromic cleft lip and palate. J. Med. Genet. 39(5):368-369.
- Rai V (2011). Evaluation of methylenetetrahydrofolate reductase gene variant (C677T) as risk factor for bipolar disorder. Cell Mol. Biol. 57:OL1558-OL1566.
- Ray JG, Meier C, Vermeulen MJ, Wyatt PR, Cole DE (2003). Association between folic acid food fortification and congenital orofacial clefts. J. Pediatr. 143:805-807.
- Rabein K, Ulrich CM (2003). 5,10- Methylenetetrahydrofolate reductase polymorphisms and leukemia risk: a HuGE minireview. Am. J. Epidemiol. 157:571-82.
- Rouget F, Monfort C, Bahuau M, Nelva A, Herman C, Francannet C, Robert-Gnansia E, Cordier S (2005). Periconceptual folates and the prevention of orofacial clefts: Role of dietary intakes in France. Rev Epidemiol. Sante Publique. 53(4):351-360.
- Shurks M, Rist PM, Kurth T (2010). 5-HTTLPR Polymorphism in the Serotonin Transporter Gene and Migraine: A Systematic Review and Meta-Analysis. Cephalalgia 30(11):1296-1305.
- Semic-Jusufagic A, Bircan R, Çelebiler O, Erdim M, Akarsu N, Elçioğlu NH (2012). Association between C677T and A1298C MTHFR gene polymorphism and nonsyndromic orofacial clefts in the Turkish population: a case-parent study. Turk. J. Pediatr. 54:617-625.
- Shaw WC, Semb G, Nelson P, Brattström V, Mølsted K, Prahl-Andersen B, Gundlach KK (2001). The Eurocleft Project 1996–2000: Overview. J. Craniomaxillofac. Surg. 29(3):131-140.
- Shoteresuk V, Ittiwut C, Siriwan P, Angspatt A (2003). Maternal 677CT/1298AC genotype of the MTHFR gene as a risk factor for cleft lip. J. Med. Genet. 40:e64.
- Sozen MA, Tolarova MM, Spritz RA (2009). The common MTHFR C677T and A1298C variants are not associated with the risk of nonsyndromic cleft lip/palate in northern Venezuela. J. Genet. Genomics 36:283-288.
- Tolarova MM, van Rooij IA, Pastor M, van der Put NM, Goldberg AC, Hol F, et al. (1998). A common mutation in the MTHFR gene is a risk factor for nonsyndromic cleft and palate anomalies. Am. J. Hum. Genet. 63:A27.
- van Rooij IA, Ocké MC, Straatman H, Zielhuis GA, Merkus HM, Steegers-Theunissen RP (2004). Periconceptual folate intake by supplement and food reduces the risk of nonsyndromic cleft lip with or without cleft palate. Prev. Med. 39:689-94.
- Vanderas AP (1987). Incidence of cleft lip, cleft palate, and cleft lip and palate among races: a review. Cleft Palate J. 24:216-225.

- Verkleij-Hagoort A, Bliek J, Sayed-Tabatabaei F, Ursem N, Steegers E, Steegers-Theunissen R (2007). Hyperhomocysteinemia and MTHFR Polymorphisms in Association With Orofacial Clefts and Congenital Heart Defects: A Meta-Analysis. Am. J. Med. Genet. 143:952-960.
- Weisberg I, Tran P, Christensen B, Sibani S, Rozen A (1998). A second genetic polymorphism in methylenetetrahydrofolate reductase (MTHFR) associated with decreased enzyme activity. Mol. Genet. Metab. 64:169-72.
- Whitehead A (2002). Meta-analysis of controlled clinical trials. Wiley, Chichester, UK.
- Wilcox AJ, Lie RT, Solvoll K, Taylor J, McConnaughey DR, Abyholm F, Vindenes H, Vollset SE, Drevon CA (2007). Folic acid supplements and risk of facial clefts: National population based case-control study. Br. Med. J. 334(7591):464.
- Wu X, Wang X, Chan Y, Jia S, Luo Y, Tang W (2013). Folate metabolizing gene polymorphisms MTHFR C677T and A1298c and risk for Down syndrome offspring: a meta-analysis. Eur. J. Obstet. Gynecol. Reprod. Biol. 167(2):154-159.
- Wu YL1, Ding XX, Sun YH, Yang HY, Chen J, Zhao X, Jiang YH, Lv XL, Wu ZQ (2013). Association between MTHFR C677T polymorphism and depression: An updated meta-analysis of 26 studies. Prog. Neuropsychopharmacol. Biol. Psychiatry 46:78-85.
- Xuan C, Xiao-Yan Bai, Ge Gao, Qin Yang, Guo-Wei He (2011). Association between polymorphism of methylenetetrahydrofolate reductase (MTHFR) C677T and risk of myocardial infarction: A metaanalysis for 8,140 cases and 10,522 controls. Arch. Med. Res. 42(8):677-85.
- Yadav S, Hasan N, Marjot T, Khan MS, Prasad K, Bentley P, et al. (2013). Detailed Analysis of Gene Polymorphisms Associated with Ischemic Stroke in South Asians. PLos one 8:e57305.
- Yazdy MM, Honein MA, Xing J (2007). Reduction in orofacial clefts following folic acid fortification of the U.S. grain supply. Birth Defects Res. 79:16-23.
- Zhang MY, Miaoa L, Li YS, Hub GY (2010). Meta-analysis of the methylenetetrahydrofolate reductase C677T polymorphism and susceptibility to Alzheimer's disease. Neurosc. Res. 68:142-150.
- Zhang T, Lou J, Zhong R, Wu J, Zou L, Sun Y, Lu X, Liu L, Miao X, Xiong G (2013). Genetic Variants in the Folate Pathway and the Risk of Neural Tube Defects: A Meta-Analysis of the Published Literature. PLos one 8(4):e59570.
- Zintzaras E (2006). C677T and A1298C methylenetetrahydrofolate reductase gene polymorphisms in schizophrenia, bipolar disorder and depression: a meta-analysis of genetic association studies. Psychiatr. Genet. 16:105-115.
- Zintzaras E (2007). Maternal gene polymorphisms involved in folate metabolism and risk of Down syndrome offspring: a meta-analysis. J. Hum. Genet. 52:943-53.
- Zintzaras E, Hadjigeorgiou GM (2005). The role of G196A polymorphism in the brain-derived neurotrophic factor gene in the cause of Parkinson's disease: a meta-analysis. J. Hum. Genet. 50:560-566.