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Full Length Research Paper

Laboratory evaluation of freshly prepared juice from garlic (*Allium sativum L*.) Liliaceae as protectants against the maize weevil, *Sitophilus zeamais* (Motsch.) [Coleoptera: Curculionidae]

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Fresh prepared garlic (*Allium sativum* L.) juice, containing the antimicrobial allicin, was evaluated as a possible grain protectant against the maize weevil, *Sitophilus zeamais* (Motsch.). Each experiment was set out in completely randomized design (CRD) with four replications and a control treatment. Adult mortality and percentage weight loss were investigated. There was an observed increase in adult mortality following days of exposure in all treatments. Statistically significant (P<0.05) reduced grain loss was also observed in all the treatments when compared with the control. The juice prepared from an indigenous Nigerian garlic cultivar (GUN) was more lethal (causing 93% adult mortality), when applied topically on the freshly emerged *S. zeamais* adults, compared to the juice prepared from a clove of garlic purchased at a supermarket in Germany (GAG). High performance liquid chromatography (HPLC) analysis indicated that the amount of allicin in GUN was 1883.2 μ g/ml while that in GAG was 3500.93 μ g/ml. This study highlights the potential of *A. sativum* containing allicin for biorational control of maize grains against *S. zeamais* infestation and damage.

Key words: Allicin, Allium sativum, biopesticide, biorational control, crop protectant, Sitophilus zeamais, stored product.

INTRODUCTION

Maize (*Zea mays L.*) or corn is a major source of dietary carbohydrate as well as the most important cereal in Sub-Saharan Africa (IITA, 2009), while the maize weevil,

Sitophilus zeamais (L.) (Coleoptera: Curculionidae), is a major pest of stored maize grain in many regions of the world including Nigeria (Adedire, 2001). Although

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Abbreviations: GUN, Umuahia main market, Nigeria; GAG, supermarket in Aachen, Germany; HPLC, high performance liquid chromatography; DADS, diallyl disulfide; *TRPA1*, transient receptor potential ankyrin-1.

synthetic insecticides have long been widely used in the control of insect pests, the indiscriminate application of synthetic products has led to various problems including toxic residues in the treated products, environmental pollution and growing resistance against insecticides by insects and pests (Huang et al., 1997). There is therefore an urgent need to continue the search for eco-friendly, cheap, sustainable and safe plant protection agents that will not contaminate food products in their use as grain protectants in storage systems for small holder farmers. Moreover, because they are often viewed as "mild" on the environment, compounds of biogenic origin are generally more positively regarded compared to substances partially or completely chemically synthesized in laboratories (Slusarenko et al., 2008), and are therefore more likely to gain wider acceptance among farmers in the long run. Crop protection agents of natural origin also have the advantage of possessing novel modes of action against insects and thus have the potential to reduce the risk of cross-resistance while offering new leads for the design of target-specific molecules (Zhou et al., 2012).

There has been a heightened interest in the last few decades in plants like garlic, which have been equipped by evolution to defend themselves against invading pathogens and pests, not only because of environmental concerns trailing the use of chemically synthesized plant protection products, but also because of farmers' and consumers' preference for organic farming strategies and produce, respectively (Nwachukwu et al., 2012; Slusarenko et al., 2008). For many of such plants, protection against pathogens and pests often comes in the form of sulphur-containing secondary metabolites synthesized following external attacks on them (Nwachukwu et al., 2012). Allicin (diallylthiosulfinate), the major antimicrobial substance in garlic, has attracted the attention of investigators because of its widely acclaimed potency. Garlic is known for its positive effect on health particularly the prevention of cardiovascular diseases and certain digestive cancers (Lalla et al., 2013). Previous studies have shown that Garlic also possess some insecticidal, fungicidal, acaricidal, nematicidal and bactericidal properties (Lalla et al., 2013). With a widespread antimicrobial activity comparable to those of common antibiotics like ampicillin (Curtis et al., 2004) and penicillin (Cavallito et al., 1944), it is hardly surprising that this compound has shown activity against some of the world's most notable plant pathogens including Phytophthora infestans and Pseudoperonospora cubensis (Portz et al., 2008).

Allicin is a phytoanticipin which means that its synthesis, from preformed precursors already present in garlic, occurs prior to any external attack or irritation, and so does not involve any expenditure of energy (Nwachukwu et al., 2012; van Etten et al., 1994). Allicin is formed as a volatile organosulfur compound following the disruption of garlic tissues either by crushing, piercing, or wounding. The substrate alliin (S-allyl-L-cysteine sulfoxide) held in the cytosol prior to tissue disruption reacts with the now liberated vacuolar enzyme, alliinase to give allicin with pyruvate and ammonia as by-products.

Although the search for and use of plant materials with grain protectant ability is not new (Parugrug and Roxas, 2008; Asawalam and Emosairue, 2006; Asawalam et al., 2006; Rajapakse, 2006; Udo, 2005; Arannilewa et al., 2002; Adedire and Ajayi, 1996; Odeyemi, 1993; Lale, 1992), given the growing role of allicin from garlic in crop protection, we decided to evaluate the efficacy of freshly prepared *Allium sativum* juice as protectants of maize grains against infestation by *S. zeamais*. While significant progress has been recorded in the use of such active agents from natural products, it is hoped that concerted efforts by stored product entomologists will lead to greater success in the biorational control of insects.

MATERIALS AND METHODS

Sitophilus zeamais culture

Adult *S. zeamais* was cultured in the laboratory at $27\pm2^{\circ}$ C, 60-65% r.h and 12 h: 12 h light: dark regime. *S. zeamais* was obtained from stocks maintained at the Crop Science Laboratory, Michael Okpara University of Agriculture, Umudike, Nigeria. The food media used was whole maize grains, purchased from Umuahia main market, Abia State Nigeria. Fifty (50) pairs of *S. zeamais* were introduced into 1 l glass jars containing 400 g weevil-susceptible maize grains. The jars were then covered with nylon mesh held in place with rubber bands. Freshly emerged adults of *S. zeamais* were subsequently used for the experiments.

Preparation and application of Allium sativum

The A. sativum (Garlic) bulbs used for the study were locally purchased from Umuahia main market, Nigeria (GUN) and from a supermarket in Aachen, Germany (GAG). The garlic juice was prepared by blending axillary buds from composite garlic bulbs using a NAKAI Japan Model 1706 Extractor. Prior to high performance liquid chromatography (HPLC) determination of allicin, the juice was introduced into a sterile 50-ml Falcon tube and centrifuged (Megafuge 1.0R; Heraeus Instruments, Osterode, Germany) at 5000 rpm (3000 g) for 10 min to separate the majority of the pulp from the liquid. Remnants of the pulp were then carefully removed from the top of the liquid with a clean spatula. A diaphragm vacuum pump (Vacuubrand GmbH, Wertheim, Germany) was used to separate the remaining pulp from the pure liquid juice under pressure. The pure filtrate was then transferred to a second sterile Falcon tube and sealed preparatory to HPLC analysis.

Fifty gram of clean and uninfested weevil-susceptible Bende white maize variety used for the study were weighed, using an MP Citizen Electronic weighing balance, and subsequently introduced into four sterilized plastic vials. To each plastic vial, 1 ml of each garlic juice type was added and mixed thoroughly by manual agitation of the vials. A control experiment containing no garlic juice was also set up. Five pairs of adult *S. zeamais* were introduced into treated and untreated maize grains. The lids of the plastic vials were perforated in order to maintain aerobic conditions in the vials. Muslin textile

materials were used to secure the top of the plastic vials and served to ensure aeration while preventing entry or exit of insects. The contents of the plastic vials were then shaken gently for proper and uniform mixing. Each treatment was replicated four times. The samples were arranged in a completely randomized design on a laboratory table.

Mortality and damage assessment assays

The number of dead insects in each vial was counted at 7, 14, 21, 28 and 35 days after treatment to estimate mortality. Maize weevil mortality was assessed as

Maize weevil mortality = Number of dead insects/Total number of insects x 100.

Data on percentage adult weevil mortality were corrected using Abbott's formula (Abbott, 1925) thus:

$$P_{T} = \frac{P_{o} - P_{c}}{100 - P_{c}}$$

Where, P_{T_i} is the Corrected mortality (%); P_0 , is the observed mortality (%); P_{C_i} is the control mortality (%).

Weight loss was assessed by re-weighing the grains to determine percentage weight loss. Percentage weight loss was calculated following the method of the FAO (FAO, 1985) as follows

Percentage weight loss =
$$\frac{[UaN - (U + D)] \times 100}{UaN} \times \frac{100}{1}$$

Where, U is the Weight of undamaged fraction in the sample; N is the total number of grains in the sample; Ua, is the average weight of undamaged grains and D, is the weight of damaged fraction in the sample.

Contact toxicity test by topical application

The fresh garlic juice samples at 1 ml dosage were applied uniformly to the bottom of the plastic vials and the control was set up in which there was no garlic juice. Five male and five female adult weevils of about 5 days old were introduced separately into each vial. Each treatment was replicated four times and weevil mortality was recorded after 12, 24, 36 and 48 h of exposure. Insects were presumed dead if they remained immobile and did not respond to five jabs with a blunt dissecting probe after an arbitrary 5-min recovery period.

High-performance liquid chromatography (HPLC) determination of allicin in garlic juice

Determination of the amount of allicin in the garlic juice preparations was performed using a JASCO HPLC. The method used was taken from Krest and Keusgen (2002). HPLC-grade water was used to dilute freshly prepared garlic juice in the ratio of 1:10. Thereafter, 1 ml of the diluted sample was introduced into a sterile vial with the injection volume set at $20 \,\mu$ l.

In order to protect the column, the diluted garlic juice was passed through a polyether sulfon membrane (0.2 µm pore size, Steriflip, Millipore), before introduction into the vial and subsequent injection into the HPLC (JASCO Chromatography Data System, with intelligent UV detector, Jasco Labor-u. Datentechnik GmbH, Groß-Umstadt, Germany). 1.5 ml of a 0.05 mg ml-1 solution (in methanol) of butyl-4-hydroxybenzoate was used as internal standard. Using the HPLC software ChromPass (version 1.8.6.1), a mixed gradient elution [solvent A, 30% (v/v) HPLC grade methanol adjusted to pH 2.0 with 85% (v/v) orthophosphoric acid; solvent B, 100% HPLC grade methanol] was performed. Elution spectra were recorded between 200-600 nm with detection at 254 nm for the chromatogram.

Statistical analysis

Data obtained were subjected to analysis of variance (ANOVA) and significant difference (P>0.05) means were separated by using Student Newmans Keul (SNK) test.

RESULTS

Mortality

The effect of fresh garlic juice on the mortality of *S. zeamais* is presented in Figure 1. The results obtained show that fresh GAG juice with a mortality rate of 73% and GUN at 87% mortality rate were significantly more effective in causing *S. zeamais* mortality at 28 DAT compared to the control.

Contact toxicity test by topical application

Upon exposure to adult *S. zeamais*, fresh GUN juice caused 100% mortality, while GAG juice caused 90% mortality (Figure 2) 48 h after topical application. The control led to zero mortality.

Effect on grain weight

There was significant weight loss of the control (Figure 3) when compared with the maize grains treated with garlic juice indicating the effectiveness of the juice in offering protection to the stored maize grains. While the untreated control grains lost over 8% of original weight on average, the grains treated with GAG and GUN only lost a negligible < 0.5% of average weight after 60 days.

High- performance liquid chromatography (HPLC) analysis of *A. sativum*

The HPLC chromatograms depicting the amount of allicin in the freshly prepared garlic juice is shown in Figures 4 and 5. The amount of allicin in GUN and GAG was found to be 1.88 and 3.50 mg/ml, respectively.

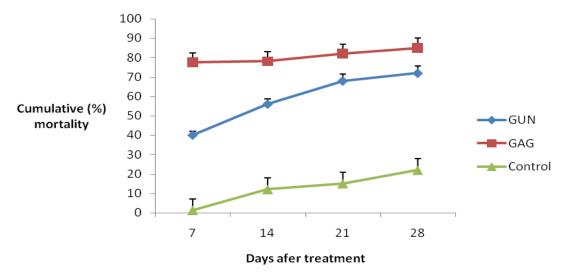


Figure 1. Cumulative mortality of freshly emerged adults of *S. zeamais* after introduction onto maize grains treated with freshly prepared *A. sativum* juice.

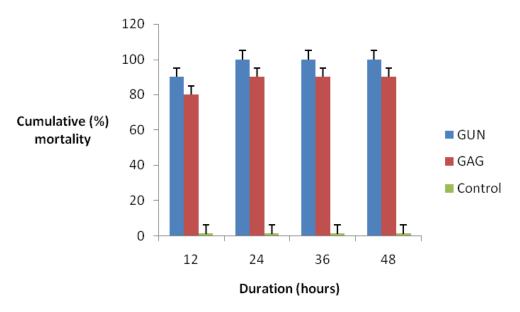


Figure 2. Contact toxicity of fresh garlic juice against S. zeamais adult at 48 h after treatment.

DISCUSSION

Over the years, various studies investigating the possible use of agents of biogenic origin as crop protectants have highlighted the potency of natural products as biopesticides (Gonzalez-Coloma et al., 2010; Lee et al., 2004; Isman, 2000; Qi and Burkholder, 1981). For instance, studies examining the fumigant and contact insecticidal effects of 22 plant essential oils against the bean weevil, *Acanthoscelides obtectus* (Regnault-Roger et al., 1993), and of 28 plant essential oils against four adult members of Order *Coleoptera* including the rice weevil *Sitophilus oryzae*, a close relative of *S. zeamais* (Shaaya et al., 1990), have not only added to the impressive body of evidence in literature clearly demonstrating the efficacy of using natural pesticides as biocontrol agents in the open field but also as stored product protectants against pests related to the maize weevil. While none of the two works just cited examined the use of agents from garlic but instead studied the use

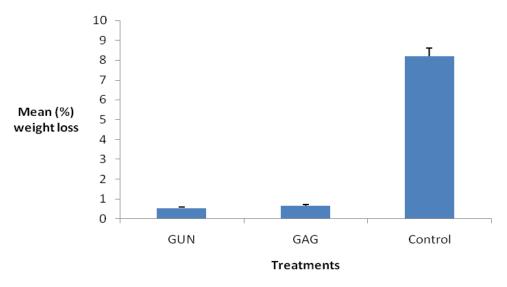


Figure 3. Effect of A. sativum juice on weight loss of maize grains.

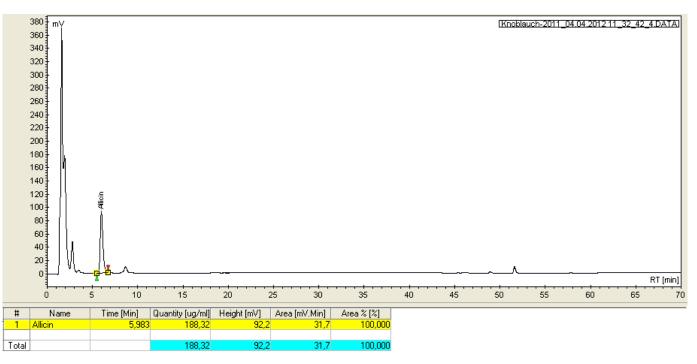
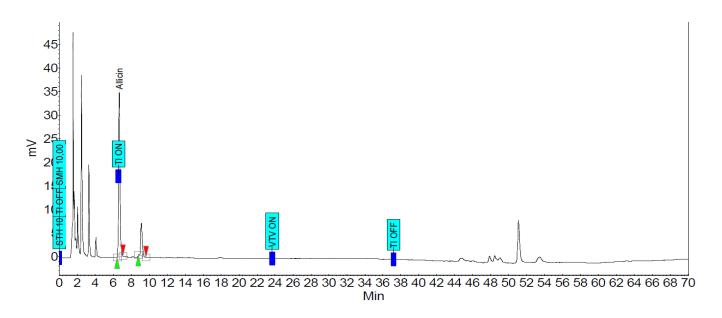


Figure 4. HPLC chromatogram for GUN allicin determination. The peak for allicin in diluted garlic juice (1:10) was detected at 5.98 min. Amount of allicin was determined to be 1883.2 µg/ml corresponding to an allicin concentration of 11.60 Mm.

of other natural compounds including those from lavender, coriander, lemon and celery, such as α -terpineol, α -terpinene, β -caryophyllen, and carvacrol among others, they provide materials for fascinating comparative analyses of chemical compounds in plants as crop protectants especially as allicin the major biologically active agent in fresh garlic juice is only found

in garlic (Cavallito and Bailey, 1944; Cavallito et al., 1944; Jain and Apitz-Castro, 1987).

Research on the potential application of biologically active compounds from garlic abound in literature. For instance, steam-distilled garlic oil has been tested for toxicity against the eggs, larvae and adults of *Tribolium castaneum* and against the adults of *S. zeamais* (Ho et



Peak results :

Index	Name	Time [Min]	Quantity [ug/ml]	Height [mV]		Area % [%]
1	Allicin	6,658	35,93	35,0	6,1	80,606
Total			35,93	41,9	7,5	100,000

Figure 5. HPLC chromatogram for GAG allicin concentration determination. The peak for allicin in diluted garlic juice (1:100) was detected at 6.66 min. The amount of allicin in the freshly prepared garlic juice was determined to be 3500.93 µg/ml corresponding to an allicin concentration of 21.57 Mm.

al., 1996) while extracts from garlic (and other plants) have been vapourized and used in fumigation tests involving T. castaneum and S. zeamais (Ho, 2000). The result from this study corroborates Musa (2013) who recorded 100% mortality at 6% w/w in groundnut seeds treated with A. sativum clove powder. Ibrahim and Garba (2011) found garlic powder to be effective in the control of maize weevil. Zhao et al (2013) observed that the essential oil of A. sativum possessed contact toxicity against overwintering Cacopsylla chinensis (Hemiptera:Psyllidae) with an LC₅₀ value of 1.42 µg per The two main constituent compounds diallyl adult. trisulfide and diallyl disulfide, exhibited strong acute toxicity against the overwintering C. Chinensis with LC₅₀ values of 0.64 and 11.04 µg per adult respectively. Feng-Lian et al. (2011) also reported that garlic essential oil diallyl disulfide and diallyl trisulfide inhibited the development of grain moth Sitotroga cerealella (Lepidoptera:Gelechidae). Ofuya et al. (2010) proved that fumigation of pods with crushed bulbs of A. sativum and Allium cepa, showed a toxic effect to Callosobruchus maculatus.

Other investigations include those of Hamed et al. (2012), Adedire and Ajayi (2006), and Arannilewa et al.

(2006). Each of these works employed garlic essential oils obtained by steam distillation or solvent extraction. However, to the best of our knowledge, no study has been carried out on the use of fresh garlic juice as a biopesticide against S. zeamais. A search of the leading electronic database, Scopus, returned no hits for each of the search terms "garlic juice weevil", "garlic juice and "garlic juice Sitophilus zeamais." Sitophilus" Therefore, this work most likely represents a remarkable shift from the conventional approach to studying the use of biologically active compounds from garlic in crop protection. Importantly, given that allicin is volatile and unstable, and upon production rapidly decomposes to breakdown products like ajoene other and the vinyldithiins (Apitz-Castro et al., 1983; Block et al., 1984; Block, 1985; Voigt and Wolf, 1986; Iberl et al., 1990), we propose that allicin could not have been principally responsible for the insecticidal effects reported in previous works investigating the exposure of agricultural pests to garlic essential oil. As comprehensively discussed by Staba et al. (2001), any of a number of treatments/processing of garlic such as freeze drying, steam distillation, oil maceration, ethanolic extraction and low temperature drying, results in the production of

complex mixtures including allicin, diallyl disulfides, dially trisulfides, allyl methyl trisulfides, ajoene, and vinyldithiins.

The results show that although GUN and GAG belong to the same species, they exert slightly different effects on *S. zeamais*- a difference which can be reasonably attributed to their different allicin contents. Our results are similar to the work of Hamed et al. (2012) which recorded mortality rates ranging from 78 -100% following 3-14 days of exposing *Sitophilus oryzae* adults to garlic essential oils. In addition to using garlic essential oils, other similar works such as those by Adedire and Ajayi (2006), and Arannilewa et al. (2006) also employed solvents like petroleum ether and ethanol as extraction vehicles thus making direct and accurate comparisons improbable.

Garlic's pungent smell has been attributed to the presence of organosulfur compounds such as allicin and diallyl disulfide (DADS) in the edible allium (Bautista et al., 2005). It has been suggested that garlic's pungency contributes to its toxicity in weevils by disrupting regular respiratory events (Adedire and Ajayi, 2006). Furthermore, the structures of allicin and DADS are similar to that of allyl isothiocyanate which apart from lending wasabi and other mustard plants their pungency, induces pain and inflammation by activating the transient receptor potential ankyrin-1 (*TRPA1*) ion channel in neuronal cells (Wang and Woolf, 2005; Jordt et al., 2004; Fahey et al., 2001).

Work with neuronal cell cultures have also provided molecular evidence suggesting that allicin is the main sulphur compounds in garlic that excites allyl isothiocyanate-sensitive sensory neurons as well as activate TRPA1 and the related TRPV1 ion channels (Bautista et al., 2005; Macpherson et al., 2005) which are present in pain-sensing neurons. Induction of pain could have significantly contributed to insect mortality by causing considerable stress the maize weevils. Finally, plant essential oils given their lipophilicity are able to penetrate the cuticle of insects (Richards, 1978) thus contributing to lethal effects.

Conclusion

The present findings suggest that freshly prepared garlic juice which has allicin as its main biologically active compound possesses a potentially vital insecticidal effect on *S. zeamais* when compared with the control. Thus, garlic offers significant promise for combating the threat posed by maize weevils to farmers in developing countries. The major thrust of this work is its adaptability for use by small scale farmers plagued by the challenge of not being able to afford conventional pesticides on the market. With no need for the more complex and sophisticated production of essential oils, this work's simplicity expressly lends zest to the overarching essence of providing a quick and easy solution to the problem of pest infestation in third world countries. There is need for further investigations to identify the other garlic juice constituents (apart from allicin) with toxic effects on *S. zeamais*, and to elucidate the precise mechanisms by which they exert their insecticidal effects.

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REFERENCES

- Abbott WS (1925). A Method of Computing the effectiveness of an insecticide. J. Econ. Entomol. 18(2):265 267.
- Adedire CO (2001) Biology, ecology and control of insect pests of stored grains. In: pp 59-94. Ofuya T.I. and Lale, N.E.S. (eds) Pest of stored cereals and pulses in Nigeria. Dave Collins Publications, Nigeria. pp. 59-94
- Adedire CO, Ajayi TS (1996). Assessment of insecticidal properties of some plants as grain protectants against the maize weevil, *Sitophilus zeamais* (Motsch.). Nig. J. Entomol. 13: 93-101.
- Apitz-Castro R, Cabrera S, Cruz MR, Ledezma E, Jain MK (1983). Effects of garlic extract and of three pure components isolated from it on human platelet aggregation, arachidonate metabolism, release reaction and platelet ultrastructure. Thromb. Res. 32(2):155-69.
- Arannilewa ST, EkrakeneT, Akinneye JO (2006). Laboratory evaluation of four medicinal plants as protectants against the maize weevil, *Sitophilus zeamais* Motsch. Afr. J. Biotechnol. 5(21):2032-2036.
- Arannilewa ST, Odeyemi OO, Adedire CO (2002). Effects of medicinal plant extract and powder on the maize weevil, *Sitophilus zeamais* Motsch. (Coleoptera: Curculionidae). Ann. Agric. Sci. 3:1-10.
- Asawalam EF, Emosairue SO (2006). Comparative efficacy of *Piper guineense* Schum and Thonn and Pirimiphos methyl on [*Sitophilus zeamais* (Motschulsky)]. Trop. Subtrop. Agroecosys. 6:143-148.
- Asawalam EF, Emosairue SO, Hassanali A (2006). Bioactivity of *Xylopia aetiopica* (Dunal) A. Rich essential oil constituents on maize weevil Sitophilus zeamais (Motschulsky). Electr. J. Environ. Agric. Food Chem. 5(1):1195-1204.
- Bautista DM, Movahed P, Hinman A, Axelsson HE, Sterner O et al. (2005). Pungent products from garlic activate the sensory ion channel TRPA1. PNAS 102(34): 12248-12252
- Block E (1985). The chemistry of garlic and onions. Sci. Am. 252(3):94-99.
- Block E, Ahmad S, Jain MK, Crecely RW, Apitz-Castro R, Cruz MR (1994). (E,Z)-ajoene: a potent antithrombotic agent from garlic. J. Am. Chem. Soc. 106(26):8295-8296.
- Cavallito CJ, Bailey JH (1944). Allicin, the antibacterial principle of *Allium sativum.* I. Isolation, physical properties and antibacterial action. J. Am. Chem. Soc. 66(11):1950-1951.
- Cavallito CJ, Buck JS, Suter CM (1944). Allicin, the antibacterial principle of *Allium sativum*. II. Determination of the chemical structure. J. Am. Chem. Soc. 66(11):1952-1954
- Curtis H, Noll U, Störmann J, Slusarenko AJ (2004). Broad-spectrum activity of the volatile phytoanticipin allicin in extracts of garlic (*Allium sativum* L.) against plant pathogenic bacteria, fungi and Oomycetes. Physiol. Mol. Plant Pathol. 65(2):79-89.
- FAO (1985). Prevention of Post-harvest food losses. Training series No.10. Italy, Rome. p. 122.

- Feng-Lian Y, Fen Z, Chao-Liang L (2011). Insecticidal activities of garlic substances against adults of grain moth, *Sitotroga cerealella* (Lepidoptera: Gelechidae) Insect Sci. 19(2): 205-212.
- Gonzalez-Coloma A, Reina M, Diaz CE, Fraga BM. (2010). Natural Product-Based Biopesticides for Insect Control. In: Comprehensive Natural Products II, Mander L, Liu HW (Eds). Elsevier. Oxford.
- IITA (2009). Cereals and legumes systems: Maize. Available at: http://old.iita.org/cms/details/maize_project_details.aspx?zoneid=63a ndarticleid=273 Accessed on May 20, 2012.
- Huang Y, Tan JMW, Kini S, Ho H (1997). Toxic and antifeedant action of nutmeg oil against *Tribolium castaneum* (Herbst) and *Sitophilus zeamais* Motsch. J. Stored Prod. Res. 33(4):289-298.
- Iberl B, Winkler G, Knobloch K (1990). Products of allicin transformation: ajoenes and dithiins, characterization and their determination by HPLC. Planta Medica 56(2): 202-211.
- Ibrahim ND, S Garba (2011). Use of garlic powder in the control of maize weevil. Proceeding of the 45 Annual conference of Agric. Soc. Nig. pp: 177-181.
- Isman MB. (2000). Plant essential oils for pest and disease management. Crop Prot. 19(8-10):603-608.
- Jain MK, Apitz-Castro R (1987). Garlic: molecular basis of the putative 'vampire-repellant' action and other matters related to heart and blood. Trends Biochem. Sci. 12:252-254.
- Jordt SE, Bautista DM, Chuang HH, McKemy DD, Zygmunt PM, Högestätt ED, Meng ID, Julius D (2004). Mustard oils and cannabinoids excite sensory nerve fibres through the TRP channel ANKTM1. Nature 427(6971):260-265.
- Krest I, Keusgen M (2002). Biosensoric flow-through method for the determination of cysteine sulfoxides. Anal. Chim. Acta. 469(2):155-164.
- Lale NES (1992). A laboratory study of the comparative toxicity of products from three spices to the maize weevil. Postharvest Biol. Technol. 2(1): 61-64.
- Lalla FD, Ahmed B, Omar A, Mohieddine M (2013) Chemical composition and biological activity of *Allium Sativum* essential oils against *Callosobruchus maculatus* J. Environ Sci. Toxicol. Food Technol. 3(1):30-36
- Lee B, Annis PC, Tumaalii F, Choi W (2004). Fumigant toxicity of essential oils from the Myrtaceae family and 1, 8-cineole against 3 major stored-grain insects. J. Stored Prod. Res. 40(5): 553-564.
- Macpherson LJ, Geierstanger BH, Viswanath V, Bandell M, Eid SR et al (2005). The pungency of garlic: activation of *TRPA1* and *TRPV1* in response to allicin. Curr. Biol. 15(10):929-934.
- Musa AK (2013). Influence of plant powders on infestation by Adults and larvae of Khapra beetle, *Trogoderma granarium* Everts (Coleoptera: Dermestidae) in stored groundnut. Aust. J. Basic Appl. Sci. 7(6):427-432.
- Nwachukwu ID, Gruhlke MCH, Slusarenko AJ (2012). Sulfur and sulfur compounds in plant defence. Nat. Prod. Commun. 7(3):395-400.
- Odeyemi OO (1993). Insecticidal properties of certain indigenous plant oils against *Sitophilus zeamais* Mots. Appl. Entomol. Phytopathol. 60(1 and 2):19-27.

- Ofuya TI, Olotuah OF, Ogunsola OJ (2010). Fumigant toxicity of crushed bulbs of two Allium to *Callosobruchus malulatus* (Fabricius) (Coleoptera: Bruchidae) Species. Chilean J. Agric. Res. 70 (3):510-514.
- Parugrug ML, Roxas AC (2008). Insecticidal action of five plants against maize weevil, *Sitophilus zeamais* Motsch. Coleoptera: Curculionidae. KMITL Sci. Technol. J. 8(1)21-38.
- Portz D, Koch E, Slusarenko AJ (2008). Effects of garlic (*Allium sativum*) juice containing allicin on *Phytophthora infestans* and downy mildew of cucumber caused by *Pseudoperonospora cubensis*. Eur. J. Plant Pathol. 122(1):197-206
- Qi IT, Burkholder WE (1981). Protection of stored wheat from the granary weevil by vegetable oils. J. Econ. Entomol. 74: 502-505.
- Rajapakse RHS (2006). The potential of plants and plant products in stored insect pest management. J. Agric. Sci. 2(1):11-21.
- Richards AG (1978). The chemistry of insect cuticle. In: Biochemistry of insects, Academic Press, New York, U.S.A. pp. 205-232.
- Slusarenko AJ, Patel A, Portz D (2008). Control of plant diseases by natural products: Allicin from garlic as a case study. Eur. J. Plant Pathol. 121(3):313-322.
- Staba John E, Lash L, Staba Joyce E (2001). A Commentary on the Effects of Garlic Extraction and Formulation on Product Composition. J. Nutr. 131: 1118S-1119S.
- Udo IO (2005). Evaluation of the potential of some local spices as stored grain protectants against the maize weevil *Sitophilus zeamais* Motsch (Coleoptera: Curculionidae). J. Appl. Sci. Environ Manage. 9 (1):165-168.
- van Etten HD, Mansfield JW, Bailey JA, Farmer EE(1994). Two classes of plant antibiotics: phytoalexins versus "phytoanticipins". Plant Cell. 6(9):1191-1192.
- Voigt M, Wolf E (1986). Knoblauch: HPLC-Bestimmung von Knoblauchwirkstoffen in Extrakten, Pulver und Fertigarzneimitteln. Dtsch. Apoth. Ztg. 126:591-593.
- Wang H, Woolf CJ (2005). Pain TRPs. Neuron 46(1):9-12.
- Zhao NN, Zhang HX, chang Liz, Zhou C, Cheng L, Qi Z, Shi W, Liv ZL, (2013) Evaluation of acute toxicity of essential of garlic (*Allium Sativum*) and it's selected major constituents against over wintering *Cacopsylla chinensis* (Hemiptera: Psyllidae.) J. Econ. Entomol. 106 (3): 1349-1354.
- Zhou HN, Zhao NN, Shu Shan D, Yang K, Cheng FW, Zhi LL, Yan JQ (2012). Insecticidal activity of the essential oil of *Lonicera japonica* flower buds and its main constituent compounds against two grain storage insects. J. Med. Plants Res. 6(5):912-917.